

Article

Cancer Simulation from Stage Minus One by Quantum microRNA Language: Lung, Colorectal and Pancreatic Cancers

Yoichi Robertus Fujii

Kawada-Cho, 106-6, Astuta-Ku, Nagoya, 456-0065, Japan;

Email: fatfujii@hotmail.co.jp; Tel.: +81-52-682-7003; Fax: +81-52-682-7003

ABSTRACT

Background: The second, third, and fourth cases of lethal cause upon cancer are lung cancer, colorectal cancer, and pancreatic cancer, respectively. This trend is not limited to some countries, but is a global problem. To further decrease cancer-related death, early prediction, diagnosis and prognosis of cancer by noninvasive liquid biopsy is an urgent issue for us. It is related with saving lives at a low medical cost and cutting-edged ideas of a neo-medical tool for risk hedge. One of the biomarkers is circulating microRNA (miRNA). miRNA can control gene expression of protein and it is a common factor of tumorigenesis and tumor suppression. Although it has recently been cleared that miRNA panel as a biomarker could be measured and evaluated as an indicator of human diseases, it is remained unclear whether the miRNA biomarker could be evaluated as biological and pathogenic processes, or pharmacologic responses to a therapeutic intervention or not.

Methods: To elucidate the implication between miRNA biomarkers and pathogenic processes, time-dependent processes of tumorigenesis in pancreatic, colorectal and lung cancers were investigated through network analysis from minus one stage (or stage zero) of cancer to various cancer stages by algorithm of miRNA entangling target sorting (METS) *in silico* simulation using quantum miRNA language.

Results: We found three different miRNA memory package (MMP) hubs for pathogenic processes among three cancer cases.

Conclusions: These computer simulation data suggested that quantum miRNA language would be essential for clinical miRNA biomarker panel to understand its biological, pathological and pharmaceutical characters in cancer.

KEYWORDS: biomarker; microRNA; colorectal cancer; lung cancer; pancreatic cancer; network; healthcare; quantum language; circRNA; lncRNA

Open Access

Received: 30 July 2019

Accepted: 21 November 2019

Published: 25 November 2019

Copyright © 2019 by the author(s). Licensee Hapres, London, United Kingdom. This is an open access article distributed under the terms and conditions of [Creative Commons Attribution 4.0 International License](https://creativecommons.org/licenses/by/4.0/).

INTRODUCTION

In USA, breast cancer (30%) in female and prostate cancer in male (19%) were estimated as the top new cases in 2017. The second and third new cases were lung cancer (12% in female and 14% in male) and colorectal cancer, respectively [1]. Pancreatic cancers were implicated in fourth estimated death (7% in both sexes) of female and male. In Japan in 2017, male lung cancer and female colorectal cancer were mentioned as the first cause of cancer-related death. Pancreatic cancer has increased in both male and female (https://ganjoho.jp/reg_stat/summary.html). These cancers have high mortality rates even when tumor cell growth is slow in general. The reason is the lack of precise and noninvasive diagnostic biomarkers before late stages of cancer. For example, pancreatic cancer is the most lethal one because most patients are diagnosed on the high stage of cancer with metastasis; however, there are some prediction tools for early detection [2]. By statistical data, the 5-year survival rate of pancreatic cancer was very low in the GLOBOCAN series (<http://globocan.iarc.fr>) of the International Agency for Research on Cancer in 2012 [3]. Although CA19-9 has been used as a biomarker of pancreatic cancer, combinations with markers of other cancer, such as CA19-9 plus CEA, CA125, CA242, or K-Ras mutation have increased sensitivity, specificity or accuracy of diagnostic screening [2,4]. It means that there would be no tumor-specific protein biomarkers, no one on one fit for quite early diagnosis. While we intend to complete precision medicine, we should challenge cancer diagnosis on minus one stage under less invasive protocols, such as a liquid biopsy. Furthermore, we should prepare treatment of regimen on the minus one stage or stage zero of cancers.

Many reports have indicated that microRNAs (miRNAs) could become promising biomarkers for quite early prediction and diagnosis of pancreatic cancer through liquid biopsy [5]. The meta-analysis from many studies showed that multiple miRNAs diagnosis has a higher sensitivity and specificity in pancreatic cancer compared with single miRNA diagnosis [6,7], as we have previously predicted in the microRNA memory package (MMP) [8,9]. Then, another meta-analysis evaluated using plasma- or serum-based miRNAs as biomarkers to diagnose pancreatic cancer [10]. They selected 27 studies from 468 papers in electric databases. It was also found that two miRNAs, miR-17-5p and miR-21 from serum exosome can distinguish pancreatic cancer from non-pancreatic cancer cases with high sensitivity of 0.73 and 0.93, and specificity of 0.96 and 0.82, respectively [11]. However, in plasma, Ganepola *et al.* [12] have showed that a panel of three miRNAs (miR-642b-3p, miR-22-3p and miR-885-5p) has higher sensitivity (0.91) and specificity (0.91) for diagnosis of early pancreatic cancer. But false-positive percentage was lowered in four miRNAs' panel (miR-1246, miR-4644, miR-3976 and miR-4306) upon exosome because the panel of miRNAs in exosome was not in healthy donors' serum exosome at all [13]. Besides exosome, to predict progression of cancer from pre-cancer state, intraductal papillary mucinous neoplasm (IPMN) is

important for early diagnosis. IPMN was pre-pancreatic cancer lesion and may develop into pancreatic cancer [14]. Three miRNAs (miR-191, miR-21 and miR-451a) in serum exosome can serve as early diagnostic and progression markers of IPMN and pancreatic cancer [15]. Furthermore, by machine learning diagnosis using two miRNA panels in plasma, pancreatic cancer could have been distinguished from chronic pancreatitis [16].

These data suggest that quantum MMP could have potential for early detection of cancer on minus one stage diagnosis. For the liquid biopsy technique, data mining and simulation would be required *in silico* as time advances, therefore, we have continued to elucidate the relation between the quantum miRNA language and the pathophysiology of human diseases. We have previously documented human breast cancer drug resistance simulation in three oncogenic subtypes by miRNA entangling target sorting (METS), in this paper, networks of pancreatic cancer, colorectal cancer and lung cancer were simulated from minus one stage or stage zero to tumor progressing stages, and their etiologies were discussed.

METHODS

Data Base Usage, Data Mining, Simulation and Statistics Tools

The physicochemical interaction has been simulated between miRNAs and human cancer diagnostic processes by dynamic computer simulation with METS algorithm using the quantum miRNA language [17]. For data mining, biomarker miRNAs were selected by following criteria: (1) data from serum or plasma, (2) statistically significant in meta-analysis, (3) showed in two or more references, (4) clear expression levels of up- and down-regulation, (5) as many between two stages in a cancer (See Table 1). Data of the multi-targets of the microRNA memory package (MMP) were extracted from miRTarBase Ver. 7.0 (<http://mirtarbase.mbc.nctu.edu.tw/php/index.php>) and TargetScan 7.2 (http://www.targetscan.org/vert_72/). In TargetScan analysis, negative correlations were observed between Context Score (CS) of miRNA/target and Double Nexus Score (DNS) as quantum energy levels of two miRNAs in all studies as described previously [8,17]. For example, a correlation (R) between CS and DNS was -0.7613 ($p < 0.01$), $R^2 = 0.5795$ in pancreatic cancer study. Target protein/protein interaction and cluster were searched by String Ver. 11.0 (<https://string-db.org/cgi/input.pl>). The gene function of protein was detected by GeneCards (<https://www.genecards.org>) and GO enrichment analysis powered by PANTHER in Geneontology (<http://geneontology.org>). To review the validated data for miRNAs, long noncoding RNAs (lncRNAs), circular RNAs (circRNAs) and cancers, PubMed (<https://www.ncbi.nlm.nih.gov/pubmed/>) and Google Scholar (<https://scholar.google.co.jp>) were used. Total information content was 944, 3076 and 6073 in pancreatic, colorectal and lung cancer, respectively. The retrieved data was narrowed down for the usage of data mining as the open accessing sub-data of 75, 208 and 314 in pancreatic, colorectal and lung cancer, respectively.

Table 1. miRNA biomarkers related with the three tumors.

	Minus-one stage (Stage 0)	Level	SNS *	Cancer	Level	SNS
Pancreatic cancer	miR-30c-5p	up	2	miR-409-3p	down	6
	miR-212-5p	up	4	miR-1246	up	2
	miR-21-5p	up	5	miR-128-3p	up	4
	miR-21-3p	down	6	miR-21-5p	up	5
	miR-10a-5p	up	5	miR-17-5p	up	7
	miR-155-5p	up	7	miR-155-5p	down	7
	miR-106b-5p	up	6	miR-196a-5p	down	8
	miR-10b-5p	up	5	miR-744-5p	up	9
Colorectal cancer	miR-29a-5p	up	5	miR-29b-1-5p	down	8
	miR-92a-1-5p	up	9	miR-17-3p	up	7
	miR-122-5p	up	9	miR-92a-1-5p	up	9
	miR-192-5p	up	4	miR-21-5p	up	5
	miR-374a-5p	up	3	miR-221-5p	up	4
	miR-29c-5p	up	5	miR-96-5p	up	4
	miR-601	up	10	miR-601	down	10
Lung cancer	miR-124-5p	up	6	miR-324-3p	up	7
	miR-154-5p	down	6	miR-1285-5p	down	5
	miR-129-2-3p	down	2	miR-21-5p	up	5
	miR-196a-3p	down	6	miR-126-5p	down	4
	miR-1180-5p	down	7	let-7a-5p	down	8
	miR-181a-2-3p	down	4	miR-145-5p	up	4
	miR-423-5p	down	10	miR-20a-5p	up	7
	miR-25-5p	down	9	miR-223-5p	up	6

Colored grids; the miRNA memory package (MMP) hub in the core of the quantum code region (QCR). * Single nexus score as quantum energy levels of a miRNA.

The calculation of statistical significance for cancer in the METS simulation was performed by the area under the curve (AUC) in receiver operating characteristic (ROC) or the χ^2 -based Cochran's Q-test using BellCurve for Excel (Social Survey Research Information Co. Ltd., Tokyo, Japan). When searching lncRNA, LncCeRBase Database (<http://insect-genome.com/LncCeRBase/front/>) was used. Accuracy and precision to develop cancer from stage minus one (or zero) were computed by machine learning using Prediction One (Sony Network Communications Inc., Tokyo, Japan).

RESULTS AND DISCUSSION

Simulation of Pancreatic Cancer

Primary extraction of miRNA set and MMP in pancreatic cancer

Duell *et al.* [18] have reported that a panel of miRNA biomarkers in pre-diagnostic plasma showed statistically significant association with subsequent risk of pancreatic ductal adenocarcinoma (PDAC) in less than

5 years. Therefore, data mining based on this report was performed for METS computation by quantum miRNA language. Eight miRNAs (miR-21-5p, miR-10b-5p, miR-106b-5p, miR-212-5p, miR-30c-5p, miR-10a-5p, miR-21-3p and miR-155) were selected as minus one stage biomarker at first (Table 1). As post-minus one stage PDAC biomarker (stage I–II: 9.2%; stage III–IV: 75%), eight miRNAs (miR-21-5p, miR-17-5p, miR-155-5p, miR-196a-5p, miR-1246, miR-744-5p, miR-409-3p and miR-128-3p) in plasma exosome were used for analysis (Table 1) [11,13].

The common miRNAs in both stages were miR-21-5p and miR-155-5p, therefore, most of the target protein gene for them were overlapped between these two miRNAs on both stages. For METS analysis, miR-106b-5p was a miRNA of miR-17/93 family, and miR-106b-5p and miR-744-5p have less target data in databases with strong evidences of protein/protein connection. Targets of miR-10b-5p was not distinguished from those of miR-10a-5p. The MMP by eight miRNA each clearly showed difference between minus-one stage and PDAC stages of pancreatic cancer (Figure 1).

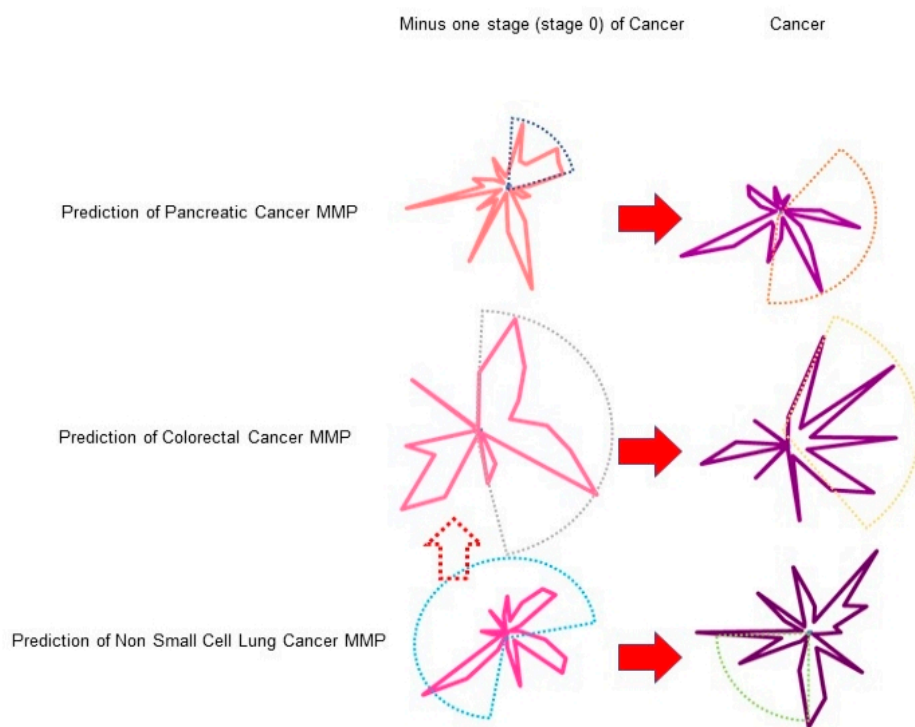


Figure 1. MMPs related with human pancreatic, colorectal and lung cancers. Three MMPs of minus one (or stage zero) stage and three MMPs of cancer stage were computed in human pancreatic, colorectal and lung cancers, and represented as radar chart. The core of the quantum code region (QCR) is shown as dotted line.

The quantum code region (QCR) in minus one stage was restricted from the energy level of 0 to 60 by DNS frequency numbers and QCR in post-minus one stage was 0 to 80 (Figure 2). To contextually elucidate difference of MMP in pancreatic cancer and quantum energy distribution of QCR between minus one and PDAC stage, etiologic causes in pancreatic cancer were dynamically computed by METS with quantum miRNA language [19].

After three layers (QCR: 0–20, 21–40 and 41–60) were learned by METS, the data on each layer were integrated into the network processing.

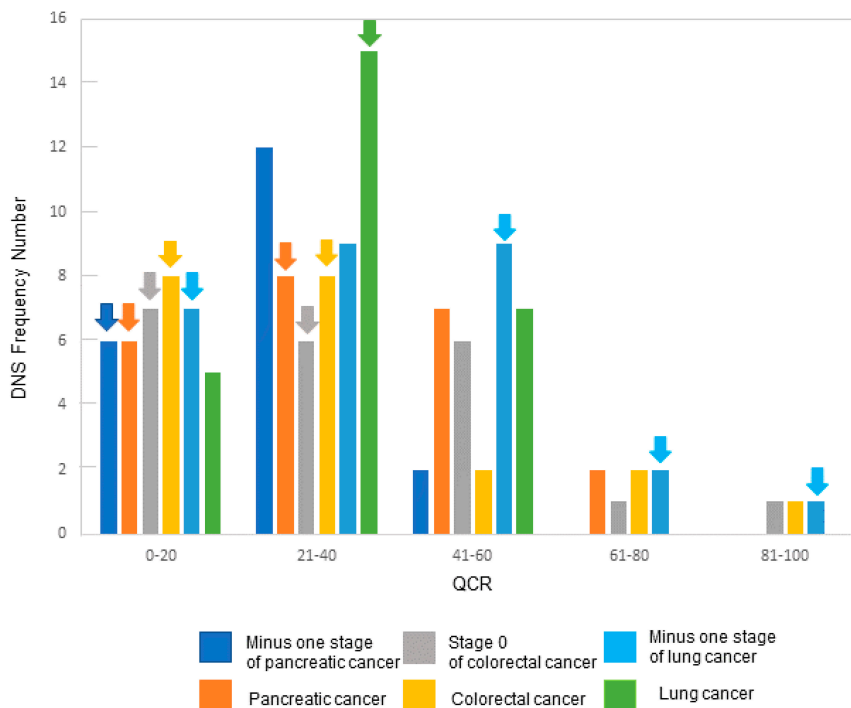


Figure 2. Alteration of DNS frequencies of human pancreatic, colorectal and lung cancers. The quantum energy level (DNS frequency) of five layers (QCR; 0–20, 21–40, 41–60, 61–80 and 81–100) was visualized in human pancreatic, colorectal and lung cancers on stages of minus one (or zero) and cancer. Arrows, the core QCR.

Minus one stage simulation of pancreatic cancer

Although a miRNA set of the minus one stage in the core stage was reduced from a panel of 7 miRNAs to MMP hub of 3 miRNAs, a main scheme of protein/protein interaction did not alter between all layers and the core (Figure 3A,B).

On minus one stage, two core miRNAs, miR-30c-5p and miR-21-5p were upregulated (Figure 3B). The upregulation of miR-30c-5p contributes to TP53 suppression together with miR-30d-5p, miR-25c-3p, miR-151a-5p, miR-612, or miR-1285-3p. BCL2 would be reduced by miR-21-5p upregulation together with miR-34a-5p, miR-34c-5p and miR-449a. The precursor of pancreatic neoplasm would be balanced in pre-tumor state between downregulation of tumor suppressor (oncogenic) and blocking of anti-apoptosis (tumor suppressive). On the contrary, miR-21-3p and miR-499a-5p are downregulated in precursor of pancreatic cancer and pancreatic cancer, respectively [18,20] and the downregulation would increase Ras-related GTP-binding protein B, RRAGB (Figure 3A,B). RRAGB activates mTOR and metabolically increases amino acids-dependent cell proliferation [21,22].

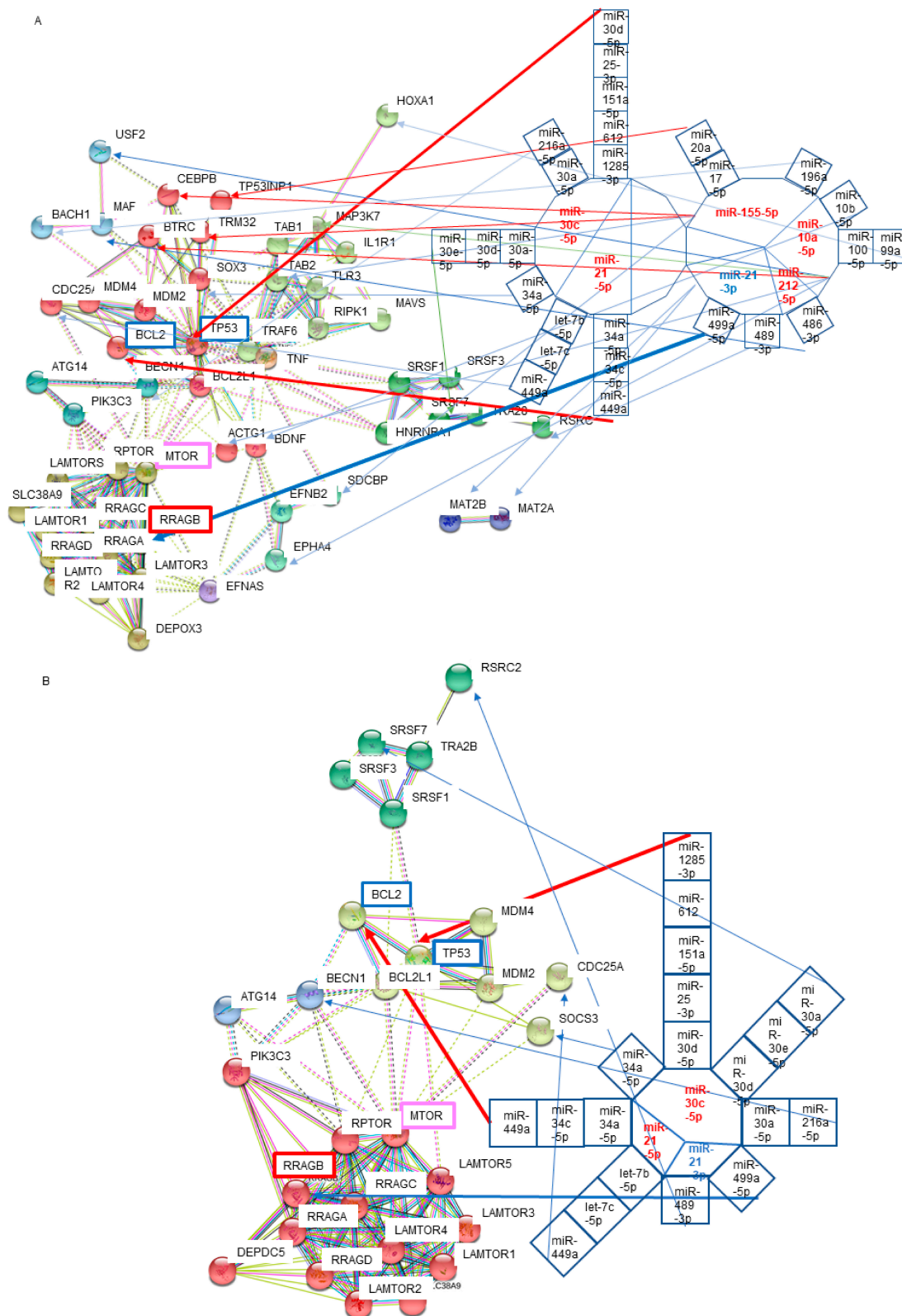


Figure 3. METS simulation of pancreatic cancer. After data mining, miRNAs of biomarker panels were selected and METS simulation was performed in pancreatic cancer on minus one stage and cancer stages. (A) Network by METS simulation and protein string clusters was represented in all layers in minus one stage of pancreatic cancer. (B) The core QCR (0–19) with MMP hub in minus one stage of pancreatic cancer. (C) All layers of PDAC. (D) The core QCR (0–39) with MMP hub of PDAC. miRNAs: upregulation—red; downregulation—blue. Proteins: augmentation—red; suppression—blue.

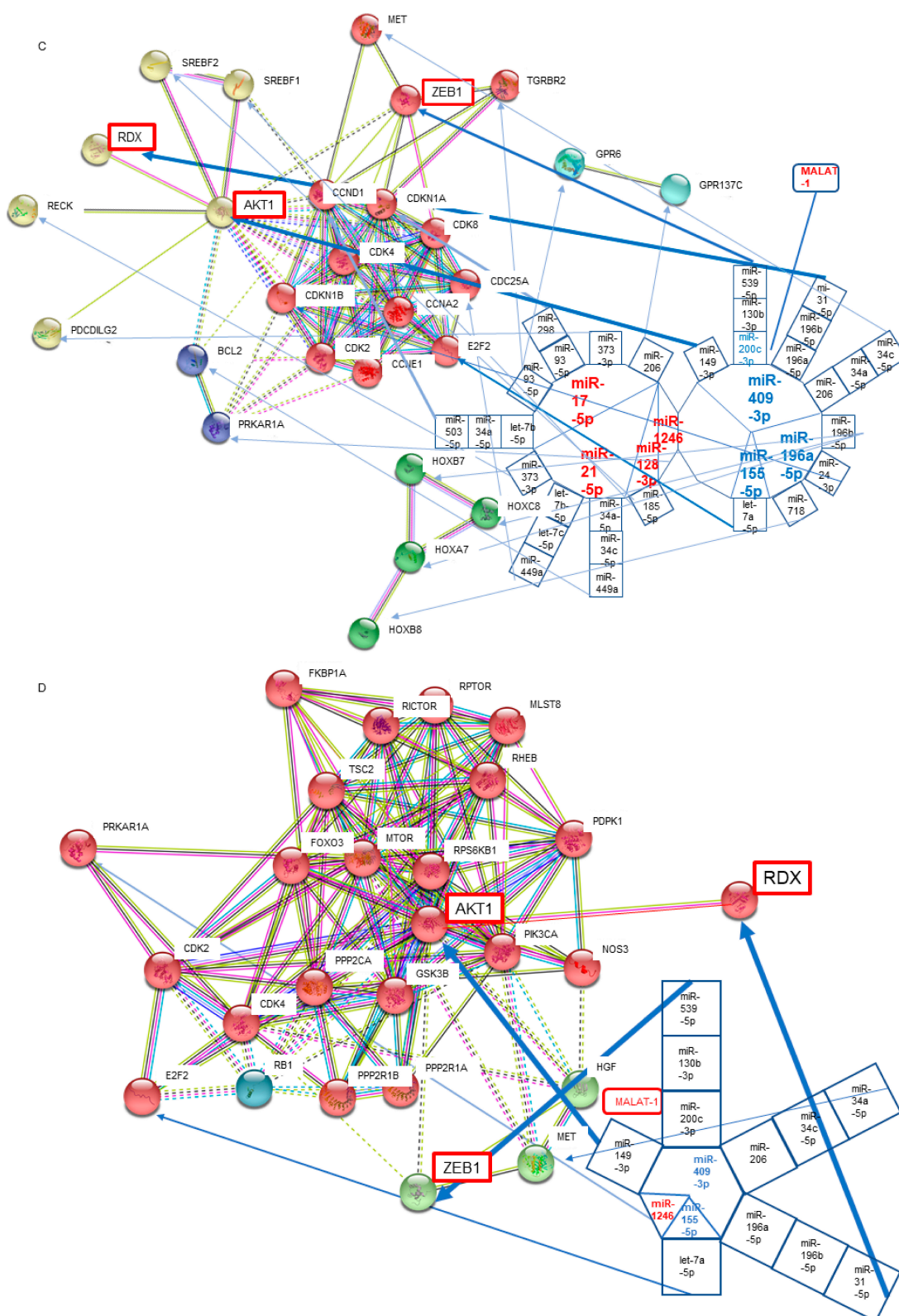


Figure 3. Cont.

mTOR/PI3K inhibitor, NVP-LED-225 inhibited pancreatic cancer stem cell [23] and everolimus showed anti-tumor effects in Panc-1 human pancreatic cancer cells by inhibition of mTOR activity through its binding [24]. Although it is known whether miR-21-3p has a role in PDAC, it is

predicted from our simulation that oncogenicity from minus one stage would be initiated by mTOR activation via miR-21-3p downregulation. While AUC in 3 MMP hub/target prediction was 0.73–0.79 ($P < 0.01$) for shorter follow up (<5 years) [18], other 4 miRNA panels would affect as tumor suppressor, such as spliceosome inhibition (target of SRSF7), cell cycle suppression (target of CDC25A), and inflammatory response inhibition (target of CEBPB). Accuracy and precision of PDAC prediction from stage minus one were 0.7813 and 0.7742, respectively. It was shown that the core QCR of minus-one stage in PDAC would be 0–19 in Figures 1, 2 and 3B.

PDAC simulation

In the case of PDAC stages in pancreatic cancer, four layers (QCR: 0–20, 21–40, 41–60, 61–80) were integrated into the network processing. Although a miRNA set of the pancreatic cancer stages in the core stage was reduced from a panel of 7 miRNAs to MMP hub of 3 miRNAs, a dominant scheme of protein/protein interaction did not alter between all layers and the core (Figure 3C,D). miR-21-5p upregulation has still inhibited BCL2, CDC25A since minus one stage (Figure 3C) and simultaneously upregulation of miR-17-5p would block CCND1, therefore, cell cycling from G1 to S would be reduced (Figure 3C). On the other hand, miR-155-5p was downregulated and E2F2 would increase. Subsequently, cell cycling would be balanced between down and up, that is shown in minus one stage because pancreatic cancer would be metastatic rather than proliferative by miR-409-3p downregulation [25,26]. miR-409-3p targeted serine-threonine kinase, AKT1 and RDX (Figure 3C,D). Upon downstream of phosphatidylinositol 3-kinase (PI3K), AKT is frequently hyperactivated in human cancer [27]. In Figure 3C,D, downregulation of miR-409-3p increased AKT with downregulation of miR-149-3p. With activation of AKT1 and its downstream of mammalian target of rapamycin, mTOR pathway enhancement is implicated in pancreatic cancer formation [28,29]. PI3K signaling affects KRAS activity in PDAC [30,31] and PI3K/AKT/mTOR has recently been targeted in therapeutic treatment of PDAC [32]. Rapamycin cotreated with cisplatin suppressed the expression of PI3K, AKT and phosphorylated mTOR in pancreatic cancer [33].

Dioscin inhibited pancreatic cancer by upregulation of miR-149-3p via suppression of AKT1 pathway [34]. In the context of human breast cancer cells, miR-409-3p was a tumor suppressor via downregulating of AKT activity [35]. For human gastric cancer, miR-409-3p was downregulated, and miR-409-3p suppressed the expression of the pro-metastatic gene radixin (RDX) [36], which was targeted by miR-409-3p with miR-196a-5p, miR-196b-5p and miR-31-5p in our simulation (Figure 3D). Further, the lncRNA, metastasis-associated lung adenocarcinoma transcript 1 (MALAT-1) expression levels were upregulated in pancreatic cancer tissues [37] and zinc finger homeodomain enhancer binding protein (ZEB1) was also upregulated [38]. PI3K promotes the metastasis of

pancreatic cancer by facilitating ZEB1 [39]. High MALAT-1 plus low miR-200c-3p expression in Figure 3C,D would promote tumor metastasis via target ZEB1, on high expression [40]. The data showed that the core QCR of PDAC was 0–39 (Figures 1, 2 and 3D).

Network analysis of PDAC

Although there is no miRNA–mRNA network computing analysis from a big database showing that aetiology of PDAC would be a disorder of AKT/mTOR pathway [41–49], these data strongly support our quantum network simulation by METS in Figure 3 that pancreatic neoplasia would progress from downstream of RRAGB/mTOR via downregulation of miR-21-3p/miR-499a-5p hub to upstream of AKT/RDX/mTOR via downregulation of miR-409-3p/miR-149-3p hub upon PDAC stages, and from low quantum levels (DNS: 30) in oncogenic state to high ones (DNS: 78) in invasion and metastatic state (Figure 3B,D). Total miRNA/PDCA prediction data of PDCA stages by METS showed AUC of 0.91 ($P < 0.01$). Therefore, PDAC simulation by quantum miRNA language was statistically confirmed (Figure 3C).

Simulation of Colorectal Cancer

A definition of stage zero in colorectal cancer

The second leading causes of cancer death is colorectal cancer (CRC) in USA [1]. Approximately over one million new cases in the global area were estimated and over half a million deaths occurred in 2012 by CRC [50]. As described above, this trend has continued in 2017. It means that the mortality of CRC has remained constant even though more screening methods of CRC have been developed, such as colonoscopy, the fecal occult blood test (FOBT), stool DNA test and double contrast barium enema (DCBE) [51]. FOBT, stool DNA test and DCBE have insufficient sensitivity with high false positive and high cost, and then colonoscopy is a semi-noninvasive test with the risk of bowel perforation because about 75% of CRC risk patients are over 60 years old (<http://gco.iarc.fr/today/home>). Further, computed tomographic (CT) colonography resulted in similar detection rate of advanced adenoma to colonoscopy [52]. CRC is classified into five stages, 0–IV. Stage I of CRC is characterized by submucosal invasion, therefore, the morphological changes and mutations of TP53 are involved in the formation of the advanced adenomas on stage 0. Before that stage, small polyps, whose sizes are less than 6 mm or histologically low dysplasia or less villous components, would be the stage minus one. The CT colonography test effectively identified diminutive polyps [53]. Because the 5-year survival rate depends on the pathological stage of CRC according to the Surveillance, Epidemiology, and End Results (SEER) data (1975–2016) of the National Cancer Institute (NCI) (<https://seer.cancer.gov/data/>) [54,55], a completely noninvasive screening methods with high

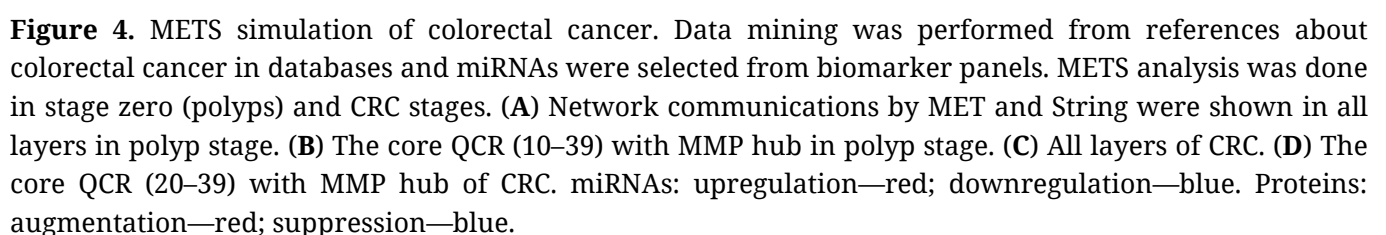
sensitivity and specificity would be required for precision medicine of CRC on early stage, minus one stage in precancerous lesions.

Primary extraction of miRNA set and MMP in CRC

Dysregulated miRNAs were identified as stage 0 of CRC in the training cohort. After data mining for METS analysis, seven miRNAs (miR-29a-5p, miR-92a-1-5p, miR-122-5p, miR-192-5p, miR-374a-5p, miR-29c-5p and miR-601) were selected as stage zero biomarker of CRC according to previous studies [56–58] (see Table 1). Seven miRNAs (miR-29b-1-5p, miR-17-3p, miR-92a-1-5p, miR-21-5p, miR-221-5p, miR-96-5p and miR-601) of the CRC stage of biomarkers from serum or plasma were also extracted from meta-analysis by Carter *et al.* [59] (see Table 1). The common miRNAs in both stages were miR-92a-1-5p and miR-601. The MMPs showed unique quantum energy levels in each stage (Figure 1) and QCRs of both stages were broad from 0 to 100 layers (Figure 2). When the data of stage zero (polyp stage) and post stage zero (CRC stages) on each layer were integrated into the network processing with METS (Figure 4A,C), the core layers were identified in QCR 10–39 of the polyp stage and QCR 20–39 of CRC stages, respectively, and interactions of protein/protein were also illustrated (Figure 4B,D).

Stage zero simulation of colorectal cancer

Although a miRNA set of the polyp stage in the core stage was reduced from a panel of 7 miRNAs to MMP hub of 3 miRNAs, a dominant scheme of protein/protein interaction did not alter between all layers and the core (Figure 4A,B). miR-192-5p plus let-7a-5p, and miR-374a-5p are upregulated and target DICER, and miR-122-5p also suppresses protein activator of interferon induced protein kinase EIF2AK2 (PRKRA) in the polyp stage (Figure 4A,B). PRKRA has similar sequences and structure to TARBP2, and TRBP2 with PACT binds PKR [60]. Both TARBP2 and PRKRA interact with Dicer and these two proteins act as cofactors of Dicer to process pre-miRNA, or could independently function as a processor distinct from Dicer pathway in mouse [61]. About implication between Dicer expression and CRC, single-nucleotide polymorphisms (SNPs) of miRNA processing gene, such as Dicer, statistically displayed a great trend of low Dicer gene expression and it would be genetically associated with CRC risk [62]. Subsequently, the SNP of rs3742330 in the human Dicer gene AA allele exhibited a significant risk of CRC, of which odds ratio is 2.11 and 95% confidence interval is from 1.33 to 3.34 ($P = 0.001$). Further, Dicer impairment induced the capacity of colorectal cancer initiation [63]. It is suggested that in stage 0, miRNA processing impairment could induce tumorigenesis of colorectal epithelial cells. Although enforced complete deletion of Dicer gene led to inhibition of tumorigenesis, partial loss of Dicer is pro-tumorigenic as described in a mouse model [64,65].



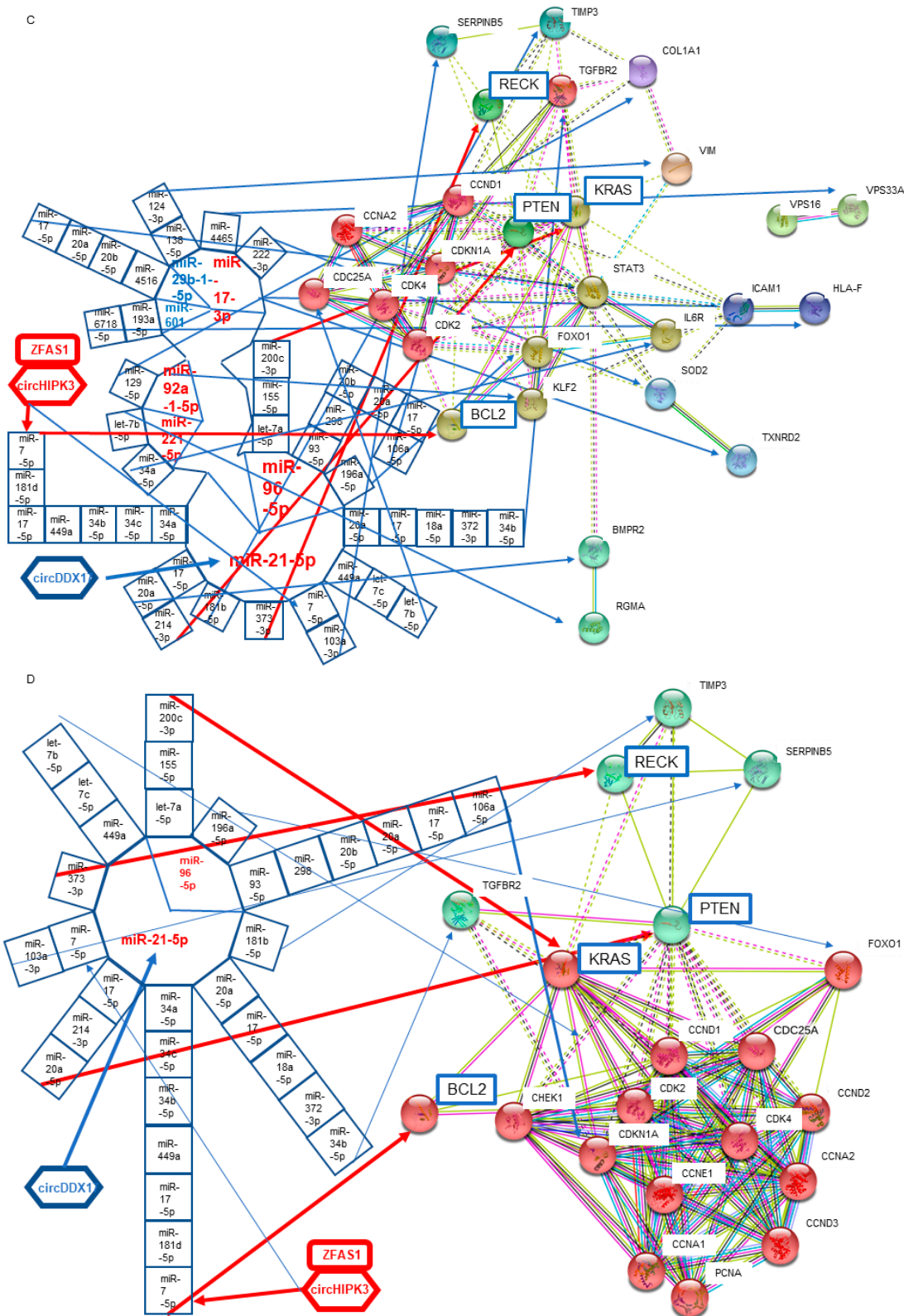


Figure 4. Cont.

Processing mechanism of human Dicer and quantum miRNA language usages in human would be different from those of mouse, therefore, species biases could lead to incorrect conclusion and speculation in bench experiments alone. However, simulation by METS could remove the

species biases which is computed by human data only. In Figure 4A, miR-192-5p with miR-34a-5p and miR-34c-5p suppressed BCL2 oncogene, and miR-122-5p with miR-630 and miR-203a-3p blocks BCL2L2 oncoprotein ligand gene. It means that early polyps may still be balanced between oncogenesis and tumor suppression. If TP53 has unfunctional mutation by dysregulation of the DNA damage repair via Dicer suppression [66,67], advanced adenoma proliferation may be progressed. Subsequent accuracy and precision of CRC prediction from stage zero (polyps) were 0.6667 and 0.7000, respectively.

CRC simulation

Although a miRNA set of the CRC stages in the core stage was reduced from a panel of 7 miRNAs to MMP hub of 2 miRNAs, a main scheme of protein/protein interaction did not alter between all layers and the core (Figure 4C,D). Core layer of CRC simulation was presented in QCR 20–39 (Figure 4D). In CRC tissues circDDX17 was significantly downregulated [68]. Therefore, upregulation of miR-21-5p by decreasing of circDDX17 suppressed PTEN tumor suppressor with miR-17-5p, miR-214-3p and miR-20a-5p, simultaneously miR-21-5p with 373-3p blocked reversion inducing cysteine rich protein with Kazal motifs (RECK), a tumor suppressor (Figure 4C,D). miR-17-5p was upregulated in CRC higher clinical stages and suppressed PTEN [69]. PTEN and RECK are deeply involved in CRC cell invasion and metastasis, and both proteins are reduced by miR-21-5p [70–72]. Furthermore, the CRC stromal cells also upregulated miR-21-5p and blocked PTEN [73].

Curcumol suppressed proliferation of CRC cells via downregulation of miR-21-5p with enhancing PTEN expression [74]. While miRNAs in exosome from stromal cells or CRC cells are transferred to the environmental cells and modulated tumorigenicity to the incorporated cells *in vitro* and *in vivo* [17,75–77], miR-21-5p from CRC stromal cells would also be incorporated into the environmental receivers and would transform the phenotype of cells to be oncogenic. miR-96-5p inhibited KRAS with let-7a-5p, miR-155-5p and miR-200c-3p. miR-21-5p suppressed BCL2 with miR-34a-5p, miR-34c-5p, miR-34b-5p, miR-449a, miR-17-5p, miR-181d-5p and miR-7-5p. Therefore, CRC cells would continue to be anti-tumor (Figure 4B). However, the concordance rate for KRAS, BRAF and PIK3CA gene mutations is presented in over 90% of primary tumors including CRC [78]. Furthermore, lncRNA ZFAS1 and circRNA circHIPK3 were significantly upregulated in CRC and sponged miR-7 [79,80]. Therefore, BCL2 suppression would be strongly modulated by both ZFAS1 and circHIPK3 as shown in Figure 4C,D. Since BCL2 expression was associated with a better prognosis in CRC [81], suppression of oncogenes KRAS and BCL2 would become negligible (Figure 4D), and KRAS and BCL2 may outflank CRC lethality [82].

Network analysis of CRC

In the network analysis of CRC, transcriptional factor and other mRNA targets were identified as hub protein genes of CRC by using the cancer genome atlas (TCGA) database, such as MYC [83–85]. Further, integrated bioinformatic computing identified more CRC hub protein genes [84–88], such as ASPN, FGF2 and CXCR4, and CRC-linked lncRNA [89,90], such as ELFN1-AS1 and HULC. However, any Dicer- and PTEN-based narrative data have no information on CRC incidents. CRC development and metastasis could dominantly be progressing with mutations of oncogene and tumor suppressor gene from stage 0 initiated by Dicer suppression via miR-192-5p and miR-374a-5p MMP hub. And then in stage I–IV, cells would be modulated by aberration of PTEN function via circDDX17 and miR-21-5p hub (Figure 4C,D). Total prediction data of noncoding RNA (ncRNA)/CRC in CRC stages by METS showed an AUC of 0.99 ($P < 0.001$), therefore, statistically, CRC simulation by quantum miRNA language was significant (Figure 4C).

Simulation of Lung Cancer

Smoking and MMP

Lung cancer is classified into two pathological subtypes, non-small cell lung cancer (NSCLC) and small cell lung cancer (SCLC). The NSCLC accounts for over 80% of lung cancer as described in National Comprehensive Cancer Network (NCCN) guideline (http://nccn.org/professionals/physician_gls/default.aspx). Risk factors are tobacco smoking, contact with radon, asbestos and other cancer-causing agents, family heredity of lung cancer, and pulmonary fibrosis. 5-year survival rate of lung cancer is only 5% in USA according to NCI (<https://seer.cancer.gov/data/>). Early diagnosis and prediction for NSCLC is a pressing issue. The National Institute of Health (NIH) defines a biomarker “a characteristic that is objectively measured and evaluated as an indicator of normal biological processes, pathogenic processes, or pharmacologic responses to a therapeutic intervention” [91]. Therefore, integrative computing analysis of the miRNA/mRNA network is absolutely required for miRNA biomarker panels for cancer diagnosis and prognosis because miRNA annotation is an arbitrary number.

At the same time, test for blood biomarker of lung cancer needs higher specificity and sensitivity [92]. For example, autoantibodies panels performed with 93% specificity but 40% low sensitivity. Complement fragment C4d was 89% specificity but 44% low sensitivity. The circulating tumor DNA (ctDNA) was 99% sensitivity but 59% low sensitivity for lung cancer. About miRNA biomarkers for lung cancer, the miRNA signature classifier (MSC) [93] and the miR-Test [94] resulted 81% specificity 87% sensitivity, and 75% specificity 78% sensitivity, respectively. The two panel tests are undergoing prospective validation with screening trials by 16,000 high risk subjects. Further, miRNA panels corresponding to MMP has been

evaluated as biological, pathological and pharmacological processes in most of all human diseases by computer simulation with quantum miRNA language [17,19]. But ctDNA cannot be used because ctDNA could not distinguish among each disease, such as pulmonary diseases, liver diseases and colorectal diseases at once. The low-dose computed tomography (LDCT) reduced lung cancer mortality in screening of high-risk individuals compared with annual chest radiography [95]. However, in general, CT and Magnetic Resonance Imaging (MRI) imaging have some problems, such as high cost, X-ray exposure, high rates of false-positive for noninvasive disease screening and human errors. Combination of both the miRNA panel and LDCT resulted a five-fold reduction of LDCT false-positive rate to 3.7% [93].

Thus, miRNA diagnosis biomarker would be the first choice of noninvasive screening tool for lung cancer. Here, at least we could show different pathological and etiological processes between smoking and lung cancer with MMP hub from miRNA biomarker panels (Figure 5). It is suggested that MMP hub from miRNA panel biomarker could be useful and smart for lung cancer prediction and diagnosis, further maybe for prognosis and therapy as all in one strategy as described previously in fundamental research of MIRAI [17].

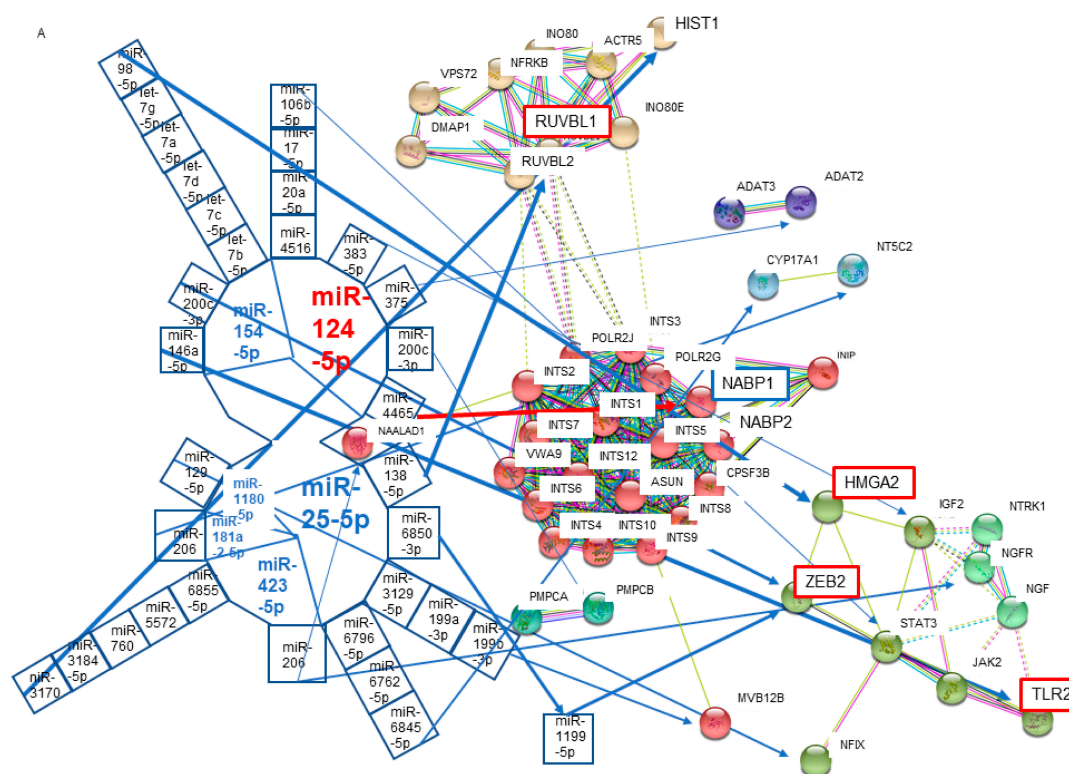


Figure 5. METS simulation of lung cancer. Data mining, and then miRNAs were selected from miRNAs biomarker panels of smoking as minus one stage and NSCLC stage I-II. Quantum data was extracted from selected miRNAs and METS analysis was performed. **(A)** Network was represented in the core QCR (0–20 and 41–90) with MMP hub of smoking. **(B)** All layers of NSCLC stage I-II. **(C)** The core QCR (31–40) with MMP hub of NSCLC stage I-II. miRNAs: upregulation—red; downregulation—blue. Proteins: augmentation—red; suppression—blue.

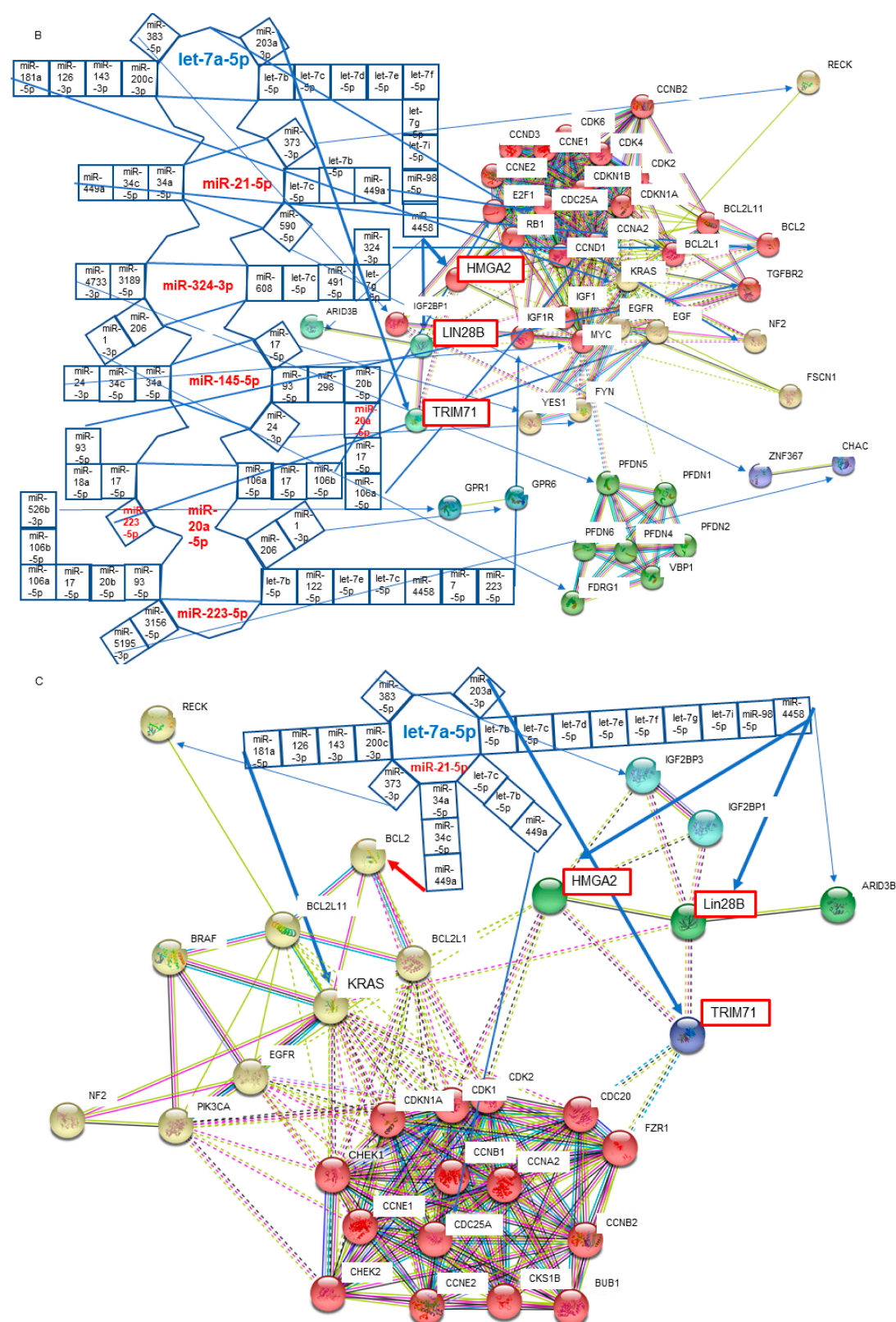


Figure 5. Cont.

Primary data extraction of lung cancer

Given NCCN clinical guideline for lung cancer listed smoking as a primary risk factor, we defined smoking as stage minus one of NSCLC. Since tobacco smoking is also listed a risk factor of CRC in NCCN Guidelines for Smoking Cessation, it would also be the stage minus one of CRC (See Figure 1). Data mining was performed for METS analysis by quantum miRNA language and then eight miRNAs (miR-124-5p, miR-154-5p, miR-129-2-3p, miR-196a-3p, miR-1180-5p, miR-181a-2-3p, miR-423-5p and miR-25-5p) were selected as biomarkers of cigarette smoking on stage minus one of lung cancer (Table 1) [96–98]. Lung cancer stage specific miRNAs have been shown in Table 2.

According to the panels, eight stage I–II specific miRNAs (miR-324-3p, miR-1285-5p, miR-21-5p, miR-126-5p, let-7a-5p, miR-145-5p, miR-20a-5p and miR-223-5p) were selected as NSCLC biomarkers for METS analysis (Table 2) [99,100]. After quantum data were extracted from the panel miRNA data, the MMPs showed unique quantum energy levels in stage minus one (smoking) and stage I–II of NSCLC (Figure 1), and QCR spectrum of smoking was from 0 to 90 of layers, and that of stage I–II in NSCLC was from 0 to 60 (Figure 2). Therefore, the core QCR spectrum of smoking was also broad in 0–20 and 41–90 but that of stage I–II is quite narrow in 31–40 (Figure 2). In smoking, miR-129-2-3p and miR-196a-3p targeted SOX4 and MYL12A, respectively. But protein/protein interaction by these two miRNAs was not significantly computed in the network simulation of all layers, therefore, all layer data of smoking showed quite similar results as described below in QCR 0–20 and 41–90 (data not shown).

Smoking simulation

Smoking miRNAs in QCR 0–20 and 41–90 were dominantly implicated in two processes, DNA repair and oncogenesis (Figure 5A).

The results as 6 miRNA MMP hub of smoking were obtained in Figure 5A (See Table 1 as well). Downregulation of miR-25-5p increased RUVBL1 with miR-138-5p (Figure 5A). RUVBL1 and RUVBL2 complex included into Pontin and Reptin complex were shown to be essential for tumorigenicity and their expression was upregulated in several cancers including lung cancer [101,102]. Upregulation of miR-124-5p represses NABP1 with miR-4465 (Figure 5A). Complex of NABP1 and NABP2 as SOSS complex promotes DNA repair on G2/M checkpoint [103]. Although carcinogens from smoking, such as nitrosamine and 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone (NNK) induces lung adenoma and carcinoma by DNA damage [104–106], in experiments using male F344 rat (species biased), the circulating and the tissue miRNA profile were significantly altered in NNK-treated group compared with control group [107,108]. Our simulation by METS also showed in human that smoking would suppress DNA repair by miR-124-5p upregulation and initiate carcinogenesis by miR-25-5p downregulation in the lung.

Table 2. Stage specific miRNAs in non-small cell lung cancer.

miRNA	Stage of NSCLC *				Specimen
	I	II	III	IV	
miR-422a	Red	Red	Red	Red	serum
miR-22	Red	Red	Red	Red	blood
miR-24	Red	Red	Red	Red	blood
miR-34a	Red	Red	Red	Red	blood
miR-125b	Red	Red	Red	Red	serum
miR-1246	Red	Red	Red	Red	serum
miR-1290	Red	Red	Red	Red	serum
let-7c	Blue	Blue	Blue	Blue	plasma
miR-152	Blue	Blue	Blue	Blue	plasma
miR-574-5p	Red	Red	Red	Red	serum
miR-874	Red	Red	Red	Red	serum
miR-21	Red	Red	Red	Red	serum
miR-126	Blue	Blue	Red	Red	serum
let-7a	Blue	Blue	Red	Red	serum
miR-125b	Red	Red	Red	Red	serum
miR-200b	Red	Red	Red	Red	serum
miR-34b	Red	Red	Red	Red	serum
miR-203	Red	Red	Red	Red	serum
miR-205	Red	Red	Red	Red	serum
miR-429	Red	Red	Red	Red	serum
miR-448	Red	Red	Red	Red	plasma
miR-4478	Red	Red	Red	Red	plasma
miR-506	Blue	Blue	Blue	Blue	plasma
miR-182	Red	Red	Red	Red	serum
miR-183	Red	Red	Red	Red	serum
miR-210	Red	Red	Red	Red	serum
miR-15b-5p	Blue	Blue	Blue	Blue	serum
miR-16-5p	Red	Red	Red	Red	serum
miR-20a-5p	Red	Red	Red	Red	serum
miR-324-3p	Red	Red	Red	Red	plasma
miR-1285	Blue	Blue	Red	Red	plasma
miR-98-5p	Red	Red	Red	Red	plasma
miR-302e	Red	Red	Red	Red	plasma
miR-495-3p	Red	Red	Red	Red	plasma
miR-613	Red	Red	Red	Red	plasma
miR-1246	Red	Red	Red	Red	serum
miR-1290	Red	Red	Red	Red	serum
miR-343-3p	Red	Red	Red	Red	plasma
miR-145-5p	Red	Red	Red	Red	plasma
miR-223-5p	Red	Red	Red	Red	plasma

Red—upregulation, Blue—downregulation. Data was referred from [99,100]. * Non-small cell lung cancer.

miR-154-5p low expression with miR-200c-3p would promote tumor via target transcriptional factor, zinc finger E-box binding homeobox 2 (ZEB2) on high expression (Figure 5A). This simulation result is well supported by ZEB2 oncogenic character that the expression level of ZEB1 and ZEB2 was correlated with NSCLC [109,110] and miR-154-5p regulated epithelial-mesenchymal transition (EMT) in NSCLC [111]. miR-25-5p targets miR-1199-5p transcripts, and then miR-1199-5p targets ZEB2 (Figure 5A). Diepenbruck and Christofori [112] showed that forced expression of miR-1199-5p in human untransformed cells was sufficient to block EMT and was embedded in a reciprocal regulation with ZEB1 and ZEB2 expression. However, they did not find any binding site of miR-1199-5p in the 3'-untranslated region (UTR) of ZEB2 in mouse (species biased) [113]. But in our TargetScan Human 7.2 analysis, ZEB2 in human has miR-1199-5p target site in the position 2972–2979 of ZEB2 3'-UTR by stronger context ++ score (−0.07) than in the position 28–34 ZEB1 3'-UTR (context ++ score; −0.03). Therefore, tumorigenic ZEB2 would be flexibly controlled by the Troika system (miR-145-5p, miR-25-5p and miR-1199-5p) under smoking. These data suggested that the computer network simulation with quantum miRNA data is absolutely necessary for cutting-edged human lung cancer research in the precision medicine initiative, because rodent experiments of tumorigenesis by a carcinogen contained species biases of different usage in quantum miRNA language among species [9], on the other hand, human clinical miRNA big data related lung cancer was preciously applied for METS *in silico* simulation.

High expression of high mobility group A2 (HMGA2) was implicated in transformation of lung cells, and inhibition of HMGA2 repressed the transformed phenotype of NSCLC in human cultured lung cells [114]. miR-154-5p with let-7-5p family target HMGA2 (Figure 5A). miR-154-5p suppressed NSCLC growth *in vitro* and *in vivo* [115], and from meta-analysis, let-7 low expression indicated a poor prognosis and HMGA2 expression was high in NSCLC patients [116]. It shows that HMGA2 upregulation by smoking is oncogenic for lung cells, and downregulation of miR-154-5p and let-7-5p family would initiate transformation of normal cells in the lung during long term smoking by quantum miRNA information summed up.

Further, Toll-like receptor 2 (TLR2) expression was significantly higher in idiopathic pulmonary fibrosis (IPF) compared to healthy individuals, and NSCLC [117], and IPF are pathogenically linked to cancer [118]. miR-154-5p downregulation augments the TLR2 expression with miR-146a-5p (Figure 5A). TLR2 are primarily expressed on the cell surface of monocytes and epithelial cells, and TLR2 is involved in inflammatory responses in mouse lung [119]. Excessive immune reaction by TLR2 high expression may induce uncontrolled pulmonary cell proliferation likely observed in IPF and would corroborate oncogenic processes of ZEB2 and HMGA2. The aggressive proliferation with suppressed DNA repair would also induce mutation of 3' UTR in KRAS [120], which was targeted by let-7-5p family in

Stage I–II of NSCLC (Figure 5B). Accuracy and precision of NSCLC prediction from stage zero smoking were 0.8333 and 0.8649, respectively.

Stage I–II simulation of NSCLC and network analysis

In the case of stage I–II of NSCLC, total layers data was integrated in Figure 5B. And then the data of core layer QCR 31–40 was re-extracted from that of all layers (Figure 5C). In all layer integration, miR-1285-5p and miR-126-5p targeted SCP2 and SNRPN, respectively, with high score. However, protein/protein interaction was not clearly observed in the network diagram (data not shown). Although a miRNA set of the stage I–II in the core stage of NSCLC was reduced from a panel of 6 miRNAs to MMP hub of 2 miRNAs, a cluster of PFDN5 (prefoldin subunit 5)/VBP1 (Von Hippel-Lindau binding protein; prefoldin subunit 3) interaction was removed from all layers of NSCLC to be built up in the core QCR (Figure 5B,C). Human PFDN has two subunits, PFDN5 and 3 makes α subunit of PFDN, and VBP1 (PFDN3) suppresses EMT of tumor cells [121]. However, PFDN1, a part of β subunit, increased metastatic growth of lung cancer [122]. Therefore, the implication between two subunits for tumor metastasis remained unclear.

Downregulation of let-7-5p family enhances expression of HMGA2 (Figure 5C), which is similar in the stage zero of smoking. Although Lin28B upregulation by downregulation of let-7-5p family was simulated in Figure 5B, Lin28B could further repress let-7-5p family expression via human ubiquitin ligase TRIM71 [123]. On the other hand, super-downregulation of let-7-5p would increase TRIM71 (Figure 5C) and TRIM71 overexpression opposed Lin28B-inducing transformation [124]. Since downregulation of miR-203a-3p and let-7-5p family would increase TRIM71 (Figure 5C), Lin28B could be suppressed by TRIM71. The above data showed that in pancreatic, colorectal and lung cancers, tumorigenic and tumor suppressive states are simultaneously presented in the balance. It is unknown whether these conditions are in one cell or cancer cells, or including environmental cells from this simulation by circulating miRNA biomarkers; however, it is certain that this event occurs in many cancer individuals at the same time. Thus, augmentation of tumor suppressive miRNA hub would be effective to reduce cancer-related death, and repression of tumorigenic miRNA hub could be a big tool of anti-cancer challenge. Given the therapeutic gadget of mixed miRNA hub has anti-oncogenic and tumor suppressive effects, cancer may be controlled more effectively than single miRNA function agent.

The KRAS 3'-UTR mutants increased risk of various cancers and changed target sensitivity score of let-7-5p via a complementary binding site (LCS6) [125]. However, the LCS6 variant appears not to be associated with prognosis of NSCLC, and there is marked lung cancer risk attributed to the LCS6 polymorphism [126]. Although CDC25A was overexpressed in NSCLC, this was true in 40% of tumor cells [127]. Furthermore, as described in colorectal cancer, KRAS, BCL2 and CDC25A may also outflank

etiology of NSCLC. Finally, stage I–II of NSCLC would still be balanced among oncogenesis and tumor suppression via let-7-5p family/Lin28/HMGA2/TRIM71 (Figure 5C).

Thus, these data suggested that DNA repair on minus one stage would be suppressed with smoking by NABP1 via miR-124-5p upregulation. Oncogenesis would be enhanced by increasing expression of HMGA2/ZEB2/TLR2 via downregulation of miR-154-5p plus let-7-5p family with miR-200c-3p or miR-446a-5p. NSCLC on stage I–II still would be balanced among oncogenic and tumor suppressive state through Lin28/HMGA2/TRIM71 via super-downregulation of let-7-5p family hub (Figure 5). Since total ncRNA/NSCLC prediction data of NSCLC stage I–II stages by METS showed an AUC of 0.98 ($P < 0.001$), NSCLC stage I–II simulation by quantum miRNA language was statistically significant (Figure 5B). In the integrated analysis of NSCLC, DNA replication and cell cycle pathways have been detected [128,129], and the WNT and the MAPK signaling pathways were implicated in lung cancer [130]; however, no let-7-5p family hub has been reported on the stage I–II of NSCLC *in silico* [131–133] even though lncRNA interactions were integrated [130,134].

CONCLUSIONS

It has been shown that profile of the miRNA gene expression would be altered by environmental factors such as chemicals, antibodies, nutrients, miRNAs and energies such as temperature, X-ray, UV and stresses, *etc.* We have previously simulated *in vivo* pharmaceutic events of medical agents with quantum language of miRNA by METS computation through deep layer learning [19]. Here, we showed that algorithm of the quantum miRNA language based on that of the quantum computing qubit could be time-dependently developed in cancer prediction from minus one stage of cancers. Further, MMP hub was extracted by the algorithm upon cancers from miRNA biomarker panel. The quantum computing algorithm can see the past and the future in time. Although the concept of algorithm in AI is quite similar to that of the quantum computing algorithm, the extraction process of miRNA hub is seemed to be analogous to our memory formation and creative intention, which would be processed from MMP [17]. According to quantum energy levels (layers, here) of MMP, the stage and cancer type of individuals were distinguished by coherence under the MMP hub of the core QCR layer, and then miRNA/target interaction would be presented as the network. The AUC of the integrated miRNA/target in each cancer prediction was high, however, accuracy and precision rates were not so high in cancer predictions from minus one stage by multivariable processing with deep learning (Table 3). For precision prediction of cancer from stage minus one, much more daily and personalized data would be required for the deep learning on multivariable processing.

Table 3. Cancer prediction from stage minus one (zero).

	Prediction One *		Excel **	
	Accuracy	Precision	AUC	
PDAC	0.7183	0.7742	0.91	(<i>p</i> < 0.01)
CRC	0.6667	0.7000	0.99	(<i>p</i> < 0.001)
NSCLC	0.8333	0.8649	0.98	(<i>p</i> < 0.001)

* Multivariable processing, ** miRNA/mRNA Two variable processing.

In summary, we found three pathogenic processes in the network: (1) with respect to pancreatic cancer, augmentation of RRAGB/mTOR via downregulation of miR-21-3p/miR-499a-5p on the stage minus one and then upstream of AKT/RDX/mTOR enhancement via downregulation of miR-409-3p/miR-149-3p on cancer stages, (2) DICER suppression via upregulation of miR-192-5p and miR-374a-5p on the stage zero of colorectal cancer and then on the stage I–IV cancer, aberration of PTEN function via downregulation of circDDX17 and upregulation of miR-21-5p, (3) suppression of DNA repair by decreasing NABP1 expression via miR-124-5p upregulation and oncogenesis by increasing HMGA2/ZEB2/TLR2 via downregulation of miR-154-5p plus let-7-5p family or miR-200c-3p or miR-446a-5p on stage minus one (smoking) of lung cancer and then on the stage I–II, balanced oncogenic plus tumor suppressive state through Lin28/HMGA2/TRIM71 upregulation via super-downregulation of let-7-5p family. The proof of concept was shown completely in computer simulations with quantum miRNA language for PDCA, CRC and NSCLC predictions. Although a biomarker has been defined by the NIH, a character of miRNA could be objectively measured and evaluated in a quantum as an indicator of biological processes, or pathogenic processes.

We have recently reported that Alzheimer’s disease and human breast cancer would be implicated in quantum miRNA language of MMP, and the pharmacokinetic drug response against breast cancer was involved in quantum miRNA energy levels [19]. Therefore, it is strongly supported that miRNA would be characterized by the previous simulation as a biomarker in the pharmacokinetic responses. The NIH definition for a biomarker also suggests that miRNA would be a biomarker not only for cancer but also for daily health management because natural substances, foods, drugs, supplements and environmental conditions could affect miRNA biomarker profiles. Minus one stage diagnosis of cancer by miRNA would be essential for cancer lethality to decrease in mass population. In the next developmental step, smart miRNA detection tool by perspiration would also be necessary for complete noninvasive biomarker to further challenge reduction of cancer-related death. In addition, contextual simulation by METS with quantum multi-layer integration would be required for precision medication to be developed for miRNA-based drugs derived from plants [135], which has been described previously [17]. In near future, combinational optimization of human MMP hub miRNAs for cancer would need the algorithm MIRAI to advance therapeutic

application of miRNA-based agents using a quantum processor. Thus, the quantum miRNA language might be useful for us to understand biomarker panels for cancers and to predict cancers upon minus on stage. Simultaneously, it would be essential for verification of the hub miRNAs in cancers by experiments and clinical trials.

CONFLICTS OF INTEREST

The author declares that there is no conflicts of interest.

REFERENCES

1. Siegel RL, Miller KD, Jemal A. Cancer Statistics, 2017. CA: Cancer J Clin. 2017;67:7-30. doi: 10.3322/caac.21387
2. Ge L, Pan B, Song F, Ma J, Zeraatkar D, Zhou J, et al. Comparing the diagnostic accuracy of five common tumor biomarkers and for a network meta-analysis of diagnostic test accuracy. BMJ Open. 2017;7:e018175. doi: 10.1136/bmjopen-2017-018175
3. Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. Int J Cancer. 2015;136:E359-86. doi: 10.1002/ijc.29210
4. Zhang Y, Yang J, Li H, Wu Y, Zhang H, Chen W. Tumor markers CA19-9, CA242 and CEA in the diagnosis of pancreatic cancer: a meta-analysis. Int J Clin Exp Med. 2015;8(7):11683-91.
5. Huang J, Liu J, Chen-Xiao K, Zhang X, Lee WN, Go VL, et al. Advance in microRNA as a potential biomarker for early detection of pancreatic cancer. Biomarker Res. 2016;4:20. doi: 10.1186/s40364-016-0074-3
6. Sun X, Zhou X, Zhang Y, Zhu X, Liu H. Systematic review and meta-analysis of diagnostic accuracy of miRNAs in patients with pancreatic cancer. Disease Markers. 2018;2018:6292396. doi: 10.1155/2018/6292396
7. Pei Z, Liu S-M, Huang J-T, Zhang X, Yan D, Xia Q, et al. Clinically relevant circulating microRNA profiling studies in pancreatic cancer using meta-analysis. Oncotarget. 2017;8(14):22616-24. doi: 10.18632/oncotarget.15148
8. Oson T, Yoshikawa M, Fujii YR. MicroRNA memory II: a novel scoring integration model for prediction of human disease by microRNA/microRNA quantum multi-interaction. J Adv Med Pharm Sci. 2016;5:1-18. doi: 10.9734/JAMPS/2016/22095
9. Yoshikawa M, Oson T, Fujii YR. MicroRNA memory I: the positive correlation between synergistic effects of microRNAs in cancer and a novel quantum scoring system. J Adv Med Pharm Sci. 2016;5:1-16. doi: 10.9734/JAMPS/2016/22134
10. Wei L, Yao K, Gan S, Suo Z. Clinical utilization of serum- or plasma-based miRNAs as early detection biomarkers for pancreatic cancer. Medicine. 2018;97:35. doi: 10.1097/MD.00000000000012132
11. Que R, Ding G, Chen J, Cao L. Analysis of serum exosomal microRNAs and clinicopathologic features of patients with pancreatic adenocarcinoma. World J Surg Oncol. 2013;11:219. doi: 10.1186/1477-7819-11-219

12. Ganepola GA, Rutledge P, Suman A, Yiengpruksawan A, Chang DH. Novel blood-based microRNA biomarker panel for early diagnosis of pancreatic cancer. *World J Gastrointest Oncol*. 2014;6(1):22-33. doi: 10.4251/wjgo.v6.i1.22
13. Madhavan B, Yue S, Galli U, Rana S, Gross W, Müller M, et al. Combined evaluation of a panel of protein and miRNA serum-exosome biomarkers for pancreatic cancer diagnosis increases sensitivity and specificity. *Int J Cancer*. 2015;136:2616-27. doi: 10.1002/ijc.29324
14. Kobayashi G, Fujita N, Maguchi H, Tanno S, Mizuno N, Hanada K, et al. Natural history of branch duct intraductal papillary mucinous neoplasm with mural nodules. *Pancreas*. 2014;43(4):532-8. doi: 10.1097/MPA.0000000000000080
15. Goto T, Fujiya M, Konishi H, Sasajima J, Fujibayashi S, Hayashi A, et al. An elevated expression of serum exosomal microRNA-191, -21, -451a of pancreatic neoplasm is considered to be efficient diagnostic marker. *BMC Cancer*. 2018;18:116. doi: 10.1186/s12885-018-4006-5
16. Cao Z, Liu C, Xu J, You L, Wang C, Lou W, et al. Plasma microRNA panels to diagnose pancreatic cancer: results from a multicenter study. *Oncotarget*. 2016;7(27):41575-83. doi: 10.18632/oncotarget.9491
17. Fujii YR. *The microRNA 2000: from HIV-1 to healthcare*. 1st ed. Irvine (CA, US): Scientific Research Publishing, Inc.; 2017.
18. Duell EJ, Lujan-Barooso L, Sala N, McElyea D, Overvad K, Tjonneland A, et al. Plasma microRNAs as biomarkers of pancreatic cancer risk in a prospective cohort study. *Int J Cancer*. 2017;141:905-15. doi: 10.1002/ijc.30790
19. Fujii YR. The quantum language of the microRNA gene and anti-cancer: with a dynamic computer simulation of human breast cancer drug resistance. *Integr Mol Med*. 2018;5(5):1-13. doi: 10.15761/IMM.1000346
20. Li F, Liang J, Bai L. MicroRNA-449a functions as a tumor suppressor in pancreatic cancer by the epigenetic regulation of ATDC expression. *Biomed Pharmacother*. 2018;103:782-9. doi: 10.1016/j.biopha.2018.04.01
21. Sancak Y, Bar-Peled L, Zoncu R, Markhard AL, Nada S, Sabatini DM. Regulator-Rag complex targets mTORC1 to the lysosomal surface and is necessary for its activation by amino acids. *Cell*. 2010;141(2):290-303. doi: 10.1016/j.cell.2010.02.024
22. Durán RV, Oppliger W, Robitaille AM, Heiserich L, Skendaj R, Gottlieb E, et al. Glutaminolysis activates Rag-mTORC1 signaling. *Mol Cell*. 2012;47:349-58. doi: 10.1016/j.molcel.2012.05.043
23. Sharma N, Nanta R, Sharma J, Gunewardena S, Singh KP, Shankar S, et al. PI3K/AKT/mTOR and sonic hedgehog pathways cooperate together to inhibit human pancreatic cancer stem cell characteristics and tumor growth. *Oncotarget*. 2015;6(31):32039-60. doi: 10.18632/oncotarget.5055
24. Liu L, Gong L, Zhang Y, Li N. Glycolysis in Panc-1 human pancreatic cells is inhibited by everolimus. *Exp Ther Med*. 2012;5:338-42. doi: 10.3892/etm.2012.787
25. Weng C, Dong H, Chen G, Zhai Y, Bai R, Hu H, et al. miR-409-3p inhibits HT1080 cell proliferation, vascularization and metastasis by targeting angiogenin. *Cancer Lett*. 2012;323(2):171-9. doi: 10.1016/j.canlet.2012.04.010

26. Kim K, Yoo D, Lee HS, Park SB, Kim C, Jo JH, et al. Identification of potential biomarkers for diagnosis of pancreatic and biliary tract cancers by sequencing of serum microRNAs. *BMC Med Genomics*. 2019;21(1):62. doi: 10.1186/s12920-019-0521-8
27. Hay N. The AKT-mTOR tango and its relevance to cancer. *Cancer Cell*. 2005;8:179-83. doi: 10.1016/j.ccr.2005.08008
28. Albury TM, Pandey V, Gitto SB, Dominguez L, Spinel LP, Talarchek J, et al. Constitutively active Akt1 cooperates with KRas^{G12D} to accelerate *in vivo* pancreatic tumor onset and progression. *Neoplasia*. 2015;17(2):175-82. doi: 10.1016/j.neo.2014.12.006
29. Hassan Z, Scheeweis C, Wirth M, Veltkamp C, Dantes Z, Feuerecker B, et al. MTOR inhibitor-based combination therapies for pancreatic cancer. *Br J Cancer*. 2018;118(3):366-77. doi: 10.1038/bjc.2017.421
30. Eser S, Reiff N, Messer M, Seider B, Gottschalk K, Dobler M, et al. Selective requirement of PI3K/PDK1 signaling for KRAS oncogene-driven pancreatic cell plasticity and cancer. *Cancer Cell*. 2013;23(3):406-20. doi: 10.1016/j.ccr.2013.01.023
31. Emmanouilidi A, Fyffe CA, Ferro R, Edling CE, Capone E, Sestito S, et al. Preclinical validation of 3-phosphoinositide-dependent protein kinase 1 inhibition in pancreatic cancer. *J Exp Clin Cancer Res*. 2019;38:191. doi: 10.1186/s13046-019-1191-2
32. Ebrahimi S, Hosseini M, Shahidsales S, Maftouh M, Ferns GA, Ghayour-Mobarhan M, et al. Targeting the AKT/PI3K signaling pathway as a potential therapeutic strategy for the treatment of pancreatic cancer. *Curr Med Chem*. 2017;24(13):1321-31. doi: 10.2174/0929867324666170206142658
33. Li B, Yang J, Lu Z, Liu B, Liu F. A study on the mechanism of rapamycin mediating the sensitivity of pancreatic cancer cells to cisplatin through PI3K/AKT/mTOR signaling pathway. *J BUON*. 2019;24(2):739-45.
34. Si L, Xu L, Yin L, Qi Y, Han X, Xu Y, et al. Potent effects of dioscin against pancreatic cancer via miR-149-3p-mediated inhibition of the AKT1 signaling pathway. *Br J Pharm*. 2017;174:553-68. doi: 10.1111/bph.13718
35. Zhang G, Liu Z, Xu H, Yang Q. miR-409-3p suppresses breast cancer cell growth and invasion by targeting AKT1. *Biochem Biophys Res Commun*. 2016;469(2):189-95. doi: 10.1016/j.bbrc.2015.11.099
36. Zheng B, Liang L, Huang S, Zha R, Liu L, Jia D, et al. MicroRNA-409 suppresses tumor cell invasion and metastasis by directly targeting radixin in gastric cancers. *Oncogene*. 2012;31:4509-16. doi: 10.1038/onc.2011.581
37. Jiao F, Hu H, Yuan C, Wang L, Jiang W, Jin Z, et al. Elevated expression level of long noncoding RNA MALAT-1 facilitates cell growth, migration and invasion in pancreatic cancer. *Oncol Rep*. 2014;32(6):2485-92. doi: 10.3892/or.2014.3518
38. Zinov'yeva MV, Nikolaev LG, Kondratyeva LG, Vinogradova TV, Sverdlov ED. Correlation between expression of *KLF5* and *ZEB1* transcription factor genes in pancreatic cancer. *Dokl Biochem Biophys*. 2018;481(1):219-21. doi: 10.1134/S1607672918040129

39. Peng Y-P, Zhu Y, Yin L-D, Wei J-S, Liu X-C, Zhu X-L, et al. PIK3R3 promotes metastasis of pancreatic cancer via ZEB1 induced epithelial-mesenchymal transition. *Cell Phys Biochem*. 2018;46:1930-8. doi: 10.1159/000496124
40. Zhuo M, Yuan C, Han T, Cui J, Jiao F, Wang L. A novel feedback loop between high MALAT-1 and low miR-200c-3p promotes cell migration and invasion in pancreatic ductal adenocarcinoma and is predictive of poor prognosis. *BMC Cancer*. 2018;18:1032. doi: 10.1186/s12885-018-4954-9
41. Zhu M, Xu Z, Wang K, Wang N, Li Y. microRNA and gene networks in human pancreatic cancer. *Oncol Lett*. 2013;6:1133-9. doi: 10.3892/ol.2013.1521
42. Naderi E, Mostafaei M, Pourshams A, Mohamadkhani A. Network of microRNAs-mRNAs interactions in pancreatic cancer. *BioMed Res Int*. 2014;2014:534821. doi: 10.1155/2014/534821
43. Yang J, Zheng Y. Identification of miRNA-mRNA crosstalk in pancreatic cancer by integrating transcriptome analysis. *Eur Rev Med Pharmacol Sci*. 2015;19:825-34.
44. Frampton AE, Castellano L, Colombo T, Giovannetti E, Krell J, Jacob J, et al. Integrated molecular analysis to investigate the role of microRNAs in pancreatic tumour growth and progression. *Lancet*. 2015;385(Suppl. 1);S37. doi: 10.1016/S0140-6736(15)60330-X
45. Liu PF, Jiang WH, Han YT, He LF, Zhang HL, Ren H. Integrated microRNA-mRNA analysis of pancreatic ductal adenocarcinoma. *Genet Mol Res*. 2015;14(3):10288-97. doi: 10.4238/2015.August.28.14
46. Zhou H-Q, Chen Q-C, Qiu Z-T, Tan W-L, Mo C-Q, Gao S-W. Integrative microRNA-mRNA and protein-protein interaction analysis in pancreatic neuroendocrine tumors. *Eur Rev Med Pharmacol Sci*. 2016;20:2842-52.
47. Wei D-M, Jiang M-T, Lin P, Yang H, Dan Y-W, Yu Q, et al. Potential ceRNA network involved in autophagy suppression of pancreatic cancer by chloroquine diphosphate: a study based on differentially-expressed circRNAs, lncRNAs, miRNAs and mRNAs. *Int J Oncol*. 2018;54:600-26. doi: 10.3892/ijo.2018.4660
48. Wang W, Lou W, Ding B, Yang B, Lu H, Kong Q, et al. A novel mRNA-miRNA-lncRNA competing endogenous RNA triple subnetwork associated with prognosis of pancreatic cancer. *Aging*. 2019;11(9):2610-27. doi: 10.18632/aging.101933
49. Felix TF, Lopez Lapa RM, de Carvalho M, Bertoni N, Tokar T, Oliveira RA, et al. MicroRNA modulated networks of adaptive and innate immune response in pancreatic ductal carcinoma. *PLoS One*. 2019;14(5):e0217421. doi: 10.1371/journal.pone.0217421
50. Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. *CA Cancer J Clin*. 2015;65:87-108. doi: 10.3322/caac.21262
51. Maida M, Macaluso FS, Ianiro G, Mangiola F, Sinagar E, Hold G, et al. Screening of colorectal cancer: present and future. *Exper Rev Anticancer Ther*. 2017;17:1131-46. doi: 10.1080/14737140.2017.1392243
52. Kim DH, Pickhardt PJ, Taylor AJ, Leung WK, Winter TC, Hinshaw JL, et al. CT colonography versus colonoscopy for the detection of advanced neoplasia. *N Engl J Med*. 2007;357(14):1403-12. doi: 10.1056/NEJMoa070543

53. Rickhardt PJ, Pooler BD, Mbah I, Weiss JM, Kim DH. Colorectal findings at repeat CT colonography screening after initial CT colonography screening negative for polyps larger than 5 mm. *Radiology*. 2017;282(1):139-48. doi: 10.1148/radiol.2016160582
54. Phipps AI, Passarelli MN, Chan AT, Harrison TA, Jeon J, Hutter CM, et al. Common genetic variation and survival after colorectal cancer diagnosis: a genome-wide analysis. *Carcinogenesis*. 2016;37(1):87-95. doi: 10.1093/carcin/bgv161
55. Duggan MA, Anderson WF, Altekruse S, Penberthy L, Sherman ME. The surveillance, epidemiology, and end results (SEER) program and pathology: toward strengthening the critical relationship. *Am J Surg Pathol*. 2016;40(12):e94-102. doi: 10.1097/PAS.0000000000000749
56. Carter JV, Roberts HL, Pan J, Rice JD, Burton JF, Galbraith NJ, et al. A highly predictive model for diagnosis of colorectal neoplasms using plasma microRNA: improving specificity and sensitivity. *Ann Surg*. 2016;264(4):575-84. doi: 10.1097/SLA.0000000000001873
57. Huang Z, Huang D, Ni S, Peng Z, Sheng W, Du X. Plasma microRNAs are promising novel biomarkers for early detection of colorectal cancer. *Int J Cancer*. 2010;127:118-26. doi: 10.1002/ijc.25007
58. Wang Q, Huang Z, Ni S, Xiao X, Xu Q, Wang L, et al. Plasma miR-601 and miR-760 are novel biomarkers for the early detection of colorectal cancer. *PLoS One*. 2012;7(9):e44398. doi: 10.1371/journal.pone.0044398
59. Carter JV, Galbraith NJ, Yang D, Burton JF, Walker SP, Galandiuk S. Blood-based microRNAs as biomarkers for the diagnosis of colorectal cancer: a systematic review and meta-analysis. *Br J Cancer*. 2017;116:762-74. doi: 10.1038/bjc.2017.12
60. Laraki G, Clerzius G, Daher A, Melendez-Peña C, Daniels S, Gatignol A. Interactions between the double-stranded RNA-binding proteins TRBP and PACT define the Medipal domain that mediates protein-protein interactions. *RNA Biol*. 2008;5(2):920103. doi: 10.4161/rna.5.2.6069
61. Pullagura RN, Buaas B, Gray N, Krening LC, Srivastava A, Braun RE. Functional redundancy of Dicer cofactors TRBP2 and PRKRA during murine embryogenesis does not involve miRNA biogenesis. *Genetics*. 2018;208:1513-22. doi: 10.1534/genetics.118.300791
62. Zhao Y, Du Y, Zhao S, Guo Z. Single-nucleotide polymorphisms of microRNA processing machinery genes and risk of colorectal cancer. *Onco Targets Ther*. 2015;8:421-5. doi: 10.2147/OTT.S78647
63. Iliou MS, da Silva-Diz V, Carmona FJ, Ramalho-Carvalho J, Heyn H, Villanueva A, et al. Impaired DICER 1 function promotes stemness and metastasis in colon cancer. *Oncogene*. 2014;33:4003-15. doi: 10.1038/onc.2013.398
64. Davalos V, Esteller M. Rolling the dice to discover the role of DICER in tumorigenesis. *Cancer Cell*. 2012;21:717-9. doi: 10.1016/j.ccr.2012.05.030
65. Swahari V, Nakamura A, Deshmukh M. The paradox of dicer in cancer. *Mol Cell Oncol*. 2016;3(3):e1155006. doi: 10.1080/23723556.2016.1155006

66. Francia S, Michelini F, Saxena A, Tang D, de Hoon M, Anelli V, et al. Site-specific DICER and Drosha RNA products control the DNA-damage response. *Nature*. 2012;488:231-5. doi: 10.1018/nature11179
67. Wei W, Ba Z, Gao M, Wu Y, Ma Y, Amiard S, et al. A role for small RNAs in DNA double-strand break repair. *Cell*. 2012;149:101-12. doi: 10.1016/j.cell.2012.03.002
68. Li X-N, Wang Z-J, Ye C-X, Zhao B-C, Li Z-L, Yang Y. RNA sequencing reveals the expression profiles of circRNA and indicates that circDDX17 acts as a tumor suppressor in colorectal cancer. *J Exp Clin Cancer Res*. 2018;37:325. doi: 10.1186/s13046-018-1006-x
69. Fang L, Li H, Wang L, Hu J, Jin T, Wang J, Yang BB. MicroRNA-17-5p promotes chemotherapeutic drug resistance and tumor metastasis of colorectal cancer by repressing PTEN expression. *Oncotarget*. 2014;5(10):2974-87.
70. Xiong B, Cheng Y, Ma L, Zhang C. MiR-21 regulates biological behavior through the PTEN/PI-3K/Akt signaling pathway in human colorectal cancer cells. *Int J Oncol*. 2012;42:219-28. doi: 10.3892/ijo.2012.1707
71. Yazdani Y, Farazmandfar T, Azadeh H, Zekavatian Z. The prognostic effect of PTEN expression status in colorectal cancer development and evaluation of factors affecting it: miR-21 and promoter methylation. *J Biomed Sci*. 2016;23:9. doi: 10.1186/s12929-016-0228-5
72. Wu F, Yuan G, Chen J, Wang C. Network analysis based on TCGA reveal hub genes in colon cancer. *Contemp Oncol*. 2017;21(2):136-44. doi: 10.5114/wo.2017.68622
73. Bullock MD, Pickard KM, Nielsen BS, Sayan AE, Jenei V, Mellone M, et al. Pleiotropic actions of miR-21 highlight the critical role of deregulated stromal microRNAs during colorectal cancer progression. *Cell Death Dis*. 2013;4:e684. doi: 10.1038/cddis.2013.213
74. Liu H, Wang J, Tao Y, Li X, Qin J, Bai Z, et al. Curcumin inhibits colorectal cancer proliferation by targeting miR-21 and modulated PTEN/PI3K/Akt pathways. *Life Sci*. 2019;221:354-61. doi: 10.1016/j.lfs.2019.02.049
75. Flgueroa J, Phillips LM, Shahar T, Hossain A, Gumin J, Kim H, et al. Exosomes from glioma-associated mesenchymal stem cells increase the tumorigenicity of glioma stem-like cells via transfer of miR-1587. *Cancer Res*. 2017;77(21):5808-19. doi: 10.1158/0008-5472.CAN-16-2524
76. Wang L, Yin P, Wang J, Sun Z, Zhou Y, Guan X. Delivery of mesenchymal stem cells-derived extracellular vesicles with enriched miR-185 inhibits progression of OPMD. *Artif Cells Nanomed Biotechnol*. 2019;47(1):2481-91. doi: 10.1080/21691401.2019.1623232
77. Abdouh M, Floris M, Gao Z-H, Arena V, Arena M, Arena GO. Colorectal cancer-derived extracellular vesicles induce transformation of fibroblasts into colon carcinoma cells. *J Exp Clin Cancer Res*. 2019;38:257. doi: 10.1186/s13046-019-1248-2
78. Mao C, Wu X-Y, Yang Z-Y, Threapleton DE, Yuan J-Q, Yu Y-Y, et al. Concordant analysis of *KRAS*, *BRAF*, *PIK3CA* mutations, and PTEN expression between primary colorectal cancer and matched metastasis. *Sci Rep*. 2015;5:8065. doi: 10.1038/srep08065

79. Zeng K, Chen X, Xu M, Liu X, Hu X, Sun H, et al. CircHIPK3 promotes colorectal cancer growth and metastasis by sponging miR-7. *Cell Death Dis.* 2018;9:417. doi: 10.1038/s41419-018-0454-8
80. Mo D, Liu W, Li Y, Cui W. Long non-coding RNA zinc finger antisense 1 (ZFAS1) regulates proliferation, migration, invasion, and apoptosis by targeting miR-7-5p in colorectal cancer. *Med Sci Monit.* 2019;25:5150-8. doi: 10.12659/MSM.916619
81. Kaklamanis L, Savage A, Whitehouse R, Doussis-Anagnostopoulou I, Biddolph S, Tsiotos P, et al. Bcl-2 protein expression: association with p53 and prognosis in colorectal cancer. *Br J Cancer.* 1998;77(11):1864-9.
82. Jones MF, Hara T, Lal A. KRAS cold turkey: using microRNAs to target KRAS-addicted cancer. *RNA Dis.* 2015;2(1):e539. doi: 10.14800/rd.539
83. Wang H, Luo J, Liu C, Niu H, Wang J, Liu Q, et al. Investigating microRNA and transcription factor co-regulatory networks in colorectal cancer. *BMC Bioinformatics.* 2017;18:388. doi: 10.1186/s12859-017-1796-4
84. Wu S, Wu F, Jiang Z. Identification of hub genes, key miRNAs and potential molecular mechanisms of colorectal cancer. *Oncol Rep.* 2017;38:2043-50. doi: 10.3892/or.2017.5930
85. Hou X, He X, Wang K, Hou N, Fu J, Jia G, et al. Genome-wide network-based analysis of colorectal cancer identifies novel prognostic factors and an integrative prognostic index. *Cell Phys Biochem.* 2018;49:1703-16. doi: 10.1159/000493614
86. Han B, Feng D, Yu X, Zhang Y, Liu Y, Zhou L. Identification and interaction analysis of molecular markers in colorectal cancer by integrated bioinformatics analysis. *Med Sci Monit.* 2018;24:6067-77. doi: 10.12659/MSM.910106
87. Chu S, Wang H, Yu M. A putative molecular network associated with colon cancer metastasis constructed from microarray data. *World J Surg Oncol.* 2017;15:115. doi: 10.1186/s12957-017-1181-9
88. Wang J, Yu H, Ye L, Jin L, Yu M, Lv Y. Integrated regulatory mechanisms of miRNAs and targeted genes involved in colorectal cancer. *Int J Clin Exp Pathol.* 2015;8(1):517-29.
89. Li F, Li Q, Wu X. Construction and analysis for differentially expressed long non-coding RNAs and microRNAs mediated competing endogenous RNA network in colon cancer. *PLoS One.* 2018;13(2):e0192494. doi: 10.1371/journal.pone.0192494
90. Zhang Z, Pan B, Lv S, Ji Z, Wu Q, Lang R, et al. Integrating microRNA expression profiling studies to systematically evaluate the diagnostic value of microRNAs in pancreatic cancer and validate their prognostic significance with the cancer genome atlas data. *Cell Phys Biochem.* 2018;49:678-95. doi: 10.1159/000493033
91. Califf RM. Biomarker definitions and their application. *Exp Biol Med.* 2016;243(3):213-21. doi: 10.1177/1535370217750088
92. Seijo LM, Peled N, Ajona D, Boeri M, Field JK, Sozzi G, et al. Biomarkers in lung cancer screening: achievements, promises, and challenges. *J Thorac Oncol.* 2018;14(3):343-57. doi: 10.1016/j.jtho.2018.11.023

93. Sozzi G, Boeri M, Rossi M, Verri C, Suatoni P, Bravi F, et al. Clinical utility of a plasma-based miRNA signature classifier within computed tomography lung cancer screening: a correlative MILD Trail Study. *J Clin Oncol*. 2014;32(8):768-73. doi: 10.1200/JCO.2013.50.4357
94. Montani F, Marzi MJ, Dezi F, Dama E, Carletti RM, Bonizzi G, et al. miR-Test: a blood test for lung cancer early detection. *J Natl Cancer Inst*. 2015;107(6):djv063. doi: 10.1093/jnci/djv063
95. National Lung Screening Trial Research Team; Aberle DR, Adams AM, Berg CD, Black WC, Church TR, Fagerstrom RM, et al. Reduced lung-cancer mortality with low-dose computed tomographic screening. *N Engl J Med*. 2011;365(5):395-409. doi: 10.1056/NEJMoa1102873
96. Huang J, Wu J, Li Y, Li X, Yang T, Yang Q, et al. Deregulation of serum microRNA expression is associated with cigarette smoking and lung cancer. *BioMed Res Int*. 2014;2014:364316. doi: 10.1155/2014/364431
97. Banerjee A, Waters D, Camacho OM, Minet E. Quantification of plasma microRNAs in a group of healthy smokers, ex-smokers and non-smokers and correlation to biomarkers of tobacco exposure. *Biomarkers*. 2015;20(2):123-31. doi: 10.3109/1354750X.2014.1000970
98. Willinger CM, Rong J, Tanriverdi K, Courchesne PL, Huan T, Wasserman GA, et al. MicroRNA signature of cigarette smoking and evidence for a putative causal role of microRNAs in smoking-related inflammation and target organ damage. *Circ Cardiovasc Genet*. 2017;10:e001678. doi: 10.1161/CIRCGENETICS.116.001678
99. Zhou Q, Huang S-X, Zhang F, Li S-J, Liu C, Xi Y-Y, et al. MicroRNAs: a novel potential biomarker for diagnosis and therapy in patients with non-small cell lung cancer. *Cell Prolif*. 2017;50:e12394. doi: 10.1111/cpr.12394
100. Zhang H, Mao F, Shen T, Luo Q, Ding Z, Qian L, et al. Plasma miR-145, miR-20a, miR-21 and miR-223 as novel biomarkers for screening early-stage non-small cell lung cancer. *Oncol Lett*. 2017;13:669-76. doi: 10.3892/ol.2016.5462
101. Grigoletto A, Lestienne P, Rosenbaum J. The multifaced proteins Reptin and Pontin as major players in cancer. *Biochem Biophys Acta*. 2011;1815:147-57. doi: 10.1016/j.bbcan.2010.11.002
102. Mao Y-Q, Houry WA. The role of Pontin and Reptin in cellular physiology and cancer etiology. *Front Mol Biosci*. 2017;4:58. doi: 10.3389/fmolb.2017.00058
103. Nikitaki Z, Hellweg C, Georgakilas AG, Ravanat J-L. Stress-induced DNA damage biomarkers: applications and limitations. *Front Chem*. 2015;3:35. doi: 10.3389/fchem.2015.00035
104. Hecht SS, Hoffman D. Tobacco-specific nitrosamines, an important group of carcinogens in tobacco and tobacco smoke. *Carcinogenesis*. 1988;9(6):875-84. doi: 10.1093/carcin/9.6.875
105. Lee H-L, Hsueh Y-H, Chung C-J, Pu Y-S, Chang LW, Hsieh DP, et al. Correlation between the urine profile of 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone metabolites and *N*⁷-methylguanine in urothelial carcinoma patients. *Cancer Epidemiol Biomarkers*. 2008;17(12):3390-5. doi: 10.1158/1055-9965.EPI-08-0761

106. Fujii T, Shimada K, Nakai T, Ohbayashi C. MicroRNAs in smoking-related carcinogenesis: biomarkers, functions, and therapy. *J Clin Med*. 2018;7:98. doi: 10.3390/jcm7050098
107. Kalscheuer S, Zhang X, Zeng Y, Upadhyaya P. Differential expression of microRNAs in early-stage neoplastic transformation in the lungs of F344 rats chronically treated with the tobacco carcinogen 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone. *Carcinogenesis*. 2008;29(12):2394-9. doi: 10.1093/carcin/bgn209
108. Wu JJ, Yang T, Li X, Xia Y, Zhao Y, Zou F, et al. 4-(Methylnitrosamino)-1-(3-pyridyl)-1-butanone induces circulating micrRNA deregulation in early lung carcinoma. *Biomed Environ Sci*. 2014;27(1):10-16. doi: 10.3967/bes2014.011
109. Ma DJ, Liu HS, Li SQ, Qin YZ, He J, Li L, et al. Correlation of the ZEB1 expression with the incidence and prognosis of non-small lung cancer. *Eur Rev Med Pharmacol Sci*. 2019;23(4):1528-35. doi: 10.26355/eurrev_201902_17111
110. Nourmohammadi B, Tafsiri E, Rahimi A, Nourmohammadi Z, Daneshvar Kakhaki A, Cho W, et al. Expression of miR-9 and miR-200c, ZEB1, ZEB2 and E-cadherin in non-small lung cancers in Iran. *Asian Pac J Cancer Prev*. 2019;20(6):1633-9. doi: 10.31557/APJCP.2019.20.6.1633
111. Lin X, Yang Z, Zhang P, Liu Y, Shao G. miR-154 inhibits migration and invasion of human non-small lung cancer by targeting ZEB2. *Oncol Lett*. 2016;12(1):301-6. doi: 10.3892/ol.2016.4577
112. Diepenbruck M, Christofori G. Reciprocal regulatory circuits coordinating EMT plasticity. *Cell Stress*. 2017;1(3):139-40. doi: 10.15698/cst2017.12.117
113. Diepenbruck M, Tiede S, Saxena M, Ivanek R, Kalathur RKR, Lüönd F, et al. miR-1199-5p and Zeb1 function in a double-negative feedback loop potentially coordinating EMT and tumor metastasis. *Nat Commun*. 2017;8(1):1168. doi: 10.1038/s41467-017-00197-w
114. Di Cello F, Hillion J, Hristov A, Wood LJ, Mukherjee M, Schuldenfrei A, et al. HMGA2 participates in transformation in human lung cancer. *Mol Cancer Res*. 2008;6(5):743-50. doi: 10.1158/1541-7786.MCR-07-0095
115. Lin X, Yang Z, Zhang P, Shao G. miR-154 suppresses non-small cell lung cancer growth *in vitro* and *in vivo*. *Oncol Lett*. 2016;12(1):301-6.
116. Li X-X, Di X, Cong S, Wang Y, Wang K. The role of let-7 and HMGA2 in the recurrence and development of lung cancer: a systematic review and meta-analysis. *Eur Rev Med Pharmacol Sci*. 2018;22:8353-66. doi: 10.26355/eurrev_201812_16533
117. Samara KD, Antoniou KM, Karagiannis K, Margaritopoulos G, Lasithiotaki I, Koutala E, et al. Expression profiles of Tall-like receptors in non-small cell lung cancer and idiopathic pulmonary fibrosis. *Int J Oncol*. 2012;40:1397-404. doi: 10.3892/ijo.2012.1374
118. Vancheri C. Idiopathic pulmonary fibrosis and cancer: do they really look similar? *BMC Med*. 2015;13:220. doi: 10.1186/s12916-015-0478-1
119. Lugade AA, Bogner PN, Murphy TF, Thanavala Y. The role of TLR2 and bacterial lipoprotein in enhancing airway inflammation and immunity. *Front Immun*. 2011;2:10. doi: 10.3389/fimmu.2011.00010

120. Chin LJ, Ratner E, Leng S, Zhai R, Nallur S, Babar I, et al. A SNP in a *let-7* microRNA complementary site in the *KRAS* 3'UTR increases non-small cell lung cancer. *Cancer Res.* 2008;68(20):8535-40. doi: 10.1158/0008-5472.CAN-08-2129
121. Kim JA, Choi DK, Min JS, Kang I, Kim JC, Kin S, et al. VBP1 represses cancer metastasis by enhancing HIF-1 α degradation induced by pVHL. *FEBS J.* 2017;285:115-6. doi: 10.1111/febs.14322
122. Wang D, Shi W, Tang Y, Liu Y, He K, Hu Y, et al. Prefoldin 1 promotes EMT and lung cancer progression by suppressing cyclin A expression. *Oncogene.* 2017;36:885-98. doi: 10.1038/onc.2016.257
123. Lee SH, Cho S, Kim MS, Choi K, Cho JY, Gwak HS, et al. The ubiquitin ligase human TRIM71 regulates *let-7* microRNA biogenesis via modulation of Lin28B protein. *Biochem Biophys Acta.* 2014;1839:374-86. doi: 10.1016/j.bbarm.2014.02.017
124. Yin J, Kim T-H, Park N, Shin D, Choi HI, Cho SC, et al. TRIM71 suppresses tumorigenesis via modulation of Lin28B-*let-7*-HMGA2 signaling. *Oncotarget.* 2016;7(48):79854-68. doi: 10.18632/oncotarget.13036
125. Kundu ST, Nallur S, Paranjape T, Boeke M, Weidhaas JB, Slack FJ. *KRAS* alleles: The LCS6 3'UTR variant and *KRAS* coding sequence mutations in the NCI-60 panel. *Cell Cycle.* 2012;11(2):361-6. doi: 10.4161/cc.11.2.18794
126. Nelson HH, Christensen BC, Plaza SL, Wiencke JK, Marsit CM, Kelsey KT. *KRAS* mutation, *KRAS*-LCS6 polymorphism, and non-small cell lung cancer. *Lung Cancer.* 2010;69(1):51-3. doi: 10.1016/j.lungcan.2009.09.008
127. Wu W, Fan Y-H, Kemp BL, Walsh G, Mao L. Overexpression of *cdc25A* and *cdc25B* is frequent in primary non-small cell lung cancer but is not associated with overexpression of *c-myc*. *Cancer Res.* 1998;58(18):4082-5.
128. Lazar V, Suo C, Orear C, van den Oord J, Balogh Z, Guegan J, et al. Integrated molecular portrait of non-small cell lung cancers. *BMC Med Genomics.* 2013;6:53. doi: 10.1186/1755-8794-6-53
129. Zheng M-L, Zhou N-K, Huang D-L, Luo C-H. Pathway cross-talk network strategy reveals key pathways in non-small lung cancer. *J BUON.* 2017;22(5):1252-8.
130. Jin X, Guan Y, Sheng H, Liu Y. Crosstalk in competing endogenous RNA network reveals the complex molecular mechanism underlying lung cancer. *Oncotarget.* 2017;8(53):91270-80. doi: 10.18632/oncotarget.20441
131. Tran N, Abhyankar V, Nguyen K, Weidanz J, Gao J. MicroRNA dysregulatory synergistic network: discovering microRNA dysregulatory modules across subtypes in non-small cell lung cancer. *BMC Bioinformatics.* 2018;19(Suppl. 20):504. doi:10.1186/s12859-018-2536-0.
132. Hu Y, Wang L, Gu J, Qu K, Wang Y. Identification of microRNA differentially expressed in three subtypes of non-small cell lung cancer and *in silico* functional analysis. *Oncotarget.* 2017;8(43):74554-66. doi: 10.18632/oncotarget.20218
133. Shao Y, Liang B, Long F, Jiang S-J. Diagnostic microRNA biomarker discovery for non-small cell lung cancer adenocarcinoma by integrative bioinformatics analysis. *BioMed Res Int.* 2017;2017:2563085. doi: 10.1155/2017/2563085

134. Sui J, Li Y-H, Zhang Y-Q, Li C-Y, Shen X, Yao W-Z, et al. Integrated analysis of long non-coding RNA-associated ceRNA network reveals potential lncRNA biomarkers in human lung adenocarcinoma. *Int J Oncol*. 2016;49:2023-36. doi: 10.3892/ijo.2016.3716
135. Zhang S, Sang X, Hou D, Chen J, Gu H, Zhang Y, et al. Plant-derived RNAi therapeutics: a strategic inhibitor of HBsAg. *Biomaterials*. 2019;210:83-93. doi: 10.1016/j.biomaterials.2019.04.033

How to cite this article:

Fujii YR. Cancer Simulation from Stage Minus One by Quantum microRNA Language: Lung, Colorectal and Pancreatic Cancers. *Med One*. 2019;4:e190023. <https://doi.org/10.20900/mo.20190023>